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Determination of inorganic phosphate by electroanalytical methods: A Review

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Abstract

Determination of inorganic phosphate is of very high importance in environmental and health care applications. Hence knowledge of suitable analytical techniques available for phosphate sensing for different applications becomes essential. Electrochemical methods for determining inorganic phosphate have several advantages over other common techniques, including detection selectivity, stability and relative environmental insensitivity of electroactive labels. The different electrochemical sensing strategies adopted for the determination of phosphate using selective ionophores are discussed in this review. The various sensing strategies are classified based on the electrochemical detection techniques used viz., potentiometry, voltammetry, amperometry, unconventional electrochemical methods etc., The enzymatic sensing of phosphate coupled with electrochemical detection is also included. Various electroanalytical methods available in the literature are assessed for their merits in terms of selectivity, simplicity, miniaturisation, adaptability and suitability for field measurements.

Key words: electroanalysis of phosphate, review, potentiometry, amperometry, bioelectroanalytical methods

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30 **1. Introduction**

31 The determination of phosphate species in environmental samples provides essential data for
32 monitoring the health of ecosystems, investigating biogeochemical processes and for checking
33 compliance with legislation. [1]The presence of inorganic phosphate derived from fertilizers, similarly
34 to nitrates, leads to an excessive growth (eutrophication) of aquatic plants and algae that disrupts
35 aquatic life cycles, while sodium and potassium organophosphate compounds are among the most
36 used pesticides in many intensive agricultural activities and are often found in ground waters, leading
37 to severe health problems.[2] From the perspective of biology, phosphate is one of the most important
38 electrolytes and an essential component of all living organisms. Phosphate plays an important role in
39 biological processes like synthesis of ATP, DNA, and control of pH in blood or lymph fluid. In a
40 clinical setting, phosphate level in serum is determined as part of a routine blood analysis. A
41 knowledge of phosphate level in body fluids can provide useful information about several diseases
42 such as hyperparathyroidism, Vitamin D deficiency, and Fanconi syndrome.[3]Analysis of salivary
43 phosphate is considered as a biomarker for different diagnostic tests. The concentration fluctuations of
44 salivary phosphate have been investigated as indicators of ovulation of women, uremic state, and risk
45 of development of dental caries and formation of dental calculus. [4]

46 Another field where phosphate control is assuming an increasing importance is the protection
47 of the cultural heritage. It was hypothesised that phosphate plays a major role in bio-deterioration of
48 archaeological sites caused by cyanobacterial biofilms. [5] The concentration of phosphorus varies
49 from 0.2 to 10 mg L⁻¹ (6.4×10^{-6} mol L⁻¹ to 0.3×10^{-4} mol L⁻¹) in natural and waste waters and from
50 0.2 to 50 mg kg⁻¹ (6.4×10^{-6} moles kg⁻¹ to 1.5×10^{-4} moles kg⁻¹) in soil. A maximum permissible
51 concentration of phosphate in river water is 0.32×10^{-6} mol L⁻¹ (9.8 µg L⁻¹) and ranges from 0.0143
52 to 0.143×10^{-3} mol L⁻¹ (0.4418 mg L⁻¹ to 4.418 mgL⁻¹) in wastewater. As for as a diagnostic fluid, the
53 concentration of phosphate ions in human saliva is found to vary from 5 to 14×10^{-3} mol L⁻¹ (154.5
54 mgL⁻¹ to 432.6 mgL⁻¹).[6,7] It is in the range of 0.81 to 1.45×10^{-3} mol L⁻¹ (25.029 to 43.26 mg L⁻¹)

55 PO_4^{3-} in adult human serum. [8,9] Several analytical methods are being evaluated to measure
56 phosphate in clinical, environmental, industrial, and biological samples. The objective is to develop
57 methods for better detection limits, better sensitivity, and negligible interference from real sample
58 matrix, optimum analysis cost and fast response for phosphate analysis. Different analytical methods
59 such as chromatography, optical fluorescent and colorimetric based (sensing) and electrochemical
60 methods are normally being developed. [10-22]. Electrochemical methods have several advantages
61 over the other common methods. The advantages include detection selectivity, stability and relative
62 environmental insensitivity of electroactive labels. Further, spectrophotometric methods involve the
63 addition of many reagents and extraction into organic solvent is often required. Furthermore
64 electrochemical techniques allow miniaturisation and operational simplicity which are highly
65 desirable attributes for field measurements.

66 Field-based measurements provide a versatile and indeed potentially invaluable screening
67 option for monitoring inorganic phosphate ions for ecological surveys. Interest in the use of field-
68 based measurements stems from a need to provide quick on-site assessments that could cover a
69 greater geographical spread while obviating much of the costs, time delays and loss of sample
70 integrity associated with traditional laboratory-based analysis. While a variety of colorimetric spot
71 test kits are commercially available and possess supreme portability, they can be prone to interference
72 and provide, at best, qualitative results. [14] The need for quick and quantitative field measurements
73 that can be carried out by non-expert investigators could be addressed by the use of electrochemical
74 detection methods. [10,23-25] The extrapolation of such technologies to yield a viable platform for
75 field testing of phosphate appear feasible but issues of selectivity and sensitivity must be clarified.
76 With the advent of wireless sensor networks, the idea of remote sensing is becoming popular and
77 electrochemistry can offer solutions to remote sensing of phosphate ions in environmental samples
78 [26]. Further, electrochemical methods are advantageous for biological diagnostic tests. When one
79 looks into the development of glucose sensors for diabetes management, it is understood that today,
80 the majority of the 6 billion annual assays performed by self-monitoring diabetic people are
81 electrochemical. Further, continuous amperometric monitoring of glucose is nowadays attempted

82 using implanted long term glucose monitors, systems with subcutaneous ultra filtration and micro
83 dialysis fibers coupled to externally-worn sensors and reverse-iontophoretic systems.[27]These
84 developments convey the importance of electroanalytical techniques for in vivo sensing. Considering
85 the importance of phosphate in environmental and clinical sectors, it appears worthwhile to write a
86 review comprising the electrochemical approaches available for sensing phosphate. The reviews
87 available to date mostly deal with sample collections , preservation and quality assurance issues of
88 phosphate sensing.[1,28] One comprehensive review covers the bioelectroanalytical aspects of
89 phosphate sensing.[14] Compilation of the existing electroanalytical techniques, will help the
90 researchers to analyse the pros and cons of the currently practised methods. This article will review
91 the existing electroanalytical techniques for their merits in terms of selectivity, simplicity,
92 miniaturisation, adaptability and suitability for field measurements and will address the possibility of
93 interferences from other anions.

94

95 Some of the strategies [1, 3, 6, 29-36] that have been applied to the electrochemical detection
96 of phosphate anions are summarised as follows:

97

- 98 • Extraction of the phosphate anion into an inert membrane (eg., Polyvinyl chloride (PVC)
99 membrane) by a non- redoxactive host (eg cationic polymers) followed by the detection of the
100 resulting membrane potential. This forms the basis of ion-selective electrodes (ISEs), and
101 chemically modified field-effect transistors (CHEMFETs).
- 102 • Detection of the current/potential perturbation response of a redox-active host on complex
103 formation (voltammetric/amperometric). Examples of such hosts include metallocenes/
104 porphyrins/pyrroles bound to a receptor group for phosphate or metal complexes, in which the
105 coordinated metal centre shows an unsaturated coordination environment and thus can bind
106 phosphate via classical coordination chemistry.
- 107 • Investigation of electroluminescence properties of dyes like rhodamine when they bind to
108 molybdophosphates,

- 109 • Optoelectrochemical detection based on the transmittance changes induced by the adsorption
110 of phosphate on ITO (indium-tin oxide) electrodes under constant applied potential
- 111 • Indirect sensing of phosphate by observing reduction in the catalytic current for the oxidation
112 of glucose on a catalytic electrode (eg.,(NiOH)₂/NiOOH electrodes)
- 113 • Investigation of blocking of ferrocyanide electron transfer kinetics induced by phosphate
114 anions on gold electrodes modified by self assembled monolayers of thiols.
- 115 • Phosphate sensing based on facilitated ion transfer across liquid/liquid interfaces
- 116 • Electroanalytical sensing of phosphate in the presence of enzymes sensitive to phosphate
- 117 • Mass changes associated with different concentration of phosphate during the
118 electropolymerisation of ethylenedioxythiophene monomer.

119

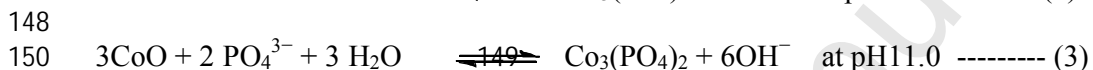
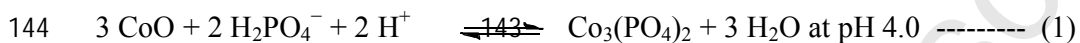
120 **2. Potentiometric detection of phosphate**

121

122 *2.1 Potentiometric Metal/Metal phosphate Sensors*

123 One of the early examples of a direct phosphate sensor was based on silver phosphate as the
124 electroactive material. Unfortunately, this ion selective electrode (ISE) method suffers from severe
125 chloride interference. [37] There are several reports on the indirect potentiometric determination of
126 phosphate using various ISEs where the activity of the electroactive cation changes because of the
127 chemical reaction of phosphate with the cation of the active material of the ISE. A cadmium ISE has
128 been used for the determination of phosphate using flow injection potentiometry (FIP). [38] This
129 method relies on detecting the decrease in $[Cd^{2+}]$ accompanying the formation of $Cd_3(PO_4)_2$ in the
130 carrier stream. This technique is susceptible to interferences from anions that also form insoluble
131 compounds with cadmium. Assaying of phosphate using continuous flow analysis in conjunction with
132 a lead ISE, [39] has also been carried out. This approach is based on detecting the ensuing decrease in
133 lead concentration following the precipitation of $Pb_3(PO_4)_2$. Unfortunately, this method also
134 experiences severe interferences due to chloride and calcium, which are among the major constituents
135 in natural waters. A ISE that has demonstrated some success for phosphate determination is based on

136 cobalt/cobalt oxide [40-44]. The response mechanism is subject to some debate, being either a host-
 137 guest relationship [40] or a mixed potential response resulting from the slow oxidation of cobalt and
 138 simultaneous reduction of both oxygen and Co^{2+} at the surface of the electrode. In addition to
 139 response to phosphate, the cobalt electrode is found to respond also to changes in the partial pressure
 140 of oxygen in the sample solution. Nevertheless, it has been shown to be capable of detecting
 141 phosphate to 0.1ppm whilst retaining a high degree of selectivity. The following reactions occurring
 142 at different pH form the basis of sensing phosphate potentiometrically.



151

152 2.2 Potentiometric studies based on organotin complexes

153 Another class of compounds used for phosphate detection is organotin complexes which respond
 154 directly to dibasic phosphate. [45] The idea of using organic tin (IV) compounds for phosphate
 155 selective electrodes was borrowed from the observation that triphenyltin compounds are good
 156 reagents for phosphate extraction.[46] Initial studies with triphenyltin dihydrogenphosphate as carrier
 157 resulted in a quite good sensitivity but poor selectivities and very slow responses .[47] Mostly,
 158 phosphate selective organotin compounds have only two organic substituents on the tin center. In
 159 contrast, trialkyltin carriers are often selective for Cl^- . Dialkyltin dinitrate ionophores were reported to
 160 give the required selectivity pattern, with a slight preference for HPO_4^{2-} over H_2PO_4^- . With the
 161 increase in the length of the alkyl chain, the interference due to other anions decreases. [48, 49] A
 162 drawback with respect to these electrodes is their positive responses towards arsenate. [49, 50] The tin
 163 (IV) centres facilitate binding of the oxygen atoms of the phosphate to the organic complex by
 164 withdrawing electrons from the tin. This electron withdrawing property, and consequent phosphate
 165 selectivity, could be further increased by replacing alkyltin compounds with benzyltin. Hence
 166 dibenzyltin dichlorides were suggested as carriers for HPO_4^{2-} .[51, 52] The investigation of
 167 dibenzyltin dichlorides with several substituents in the para position of the benzene ring indicated an

168 increase of the phosphate selectivity. [53,54] Highly hydrophilic tribasic citrate is found to interfere in
169 the case of electrodes based on bis(*p*-chlorobenzyl)tin or bis(*p*-fluorobenzyl)tin carriers . However
170 the response mechanism of these electrodes to tribasic citrate and phosphate appears to be different.
171 The severe limitation of all dibenzyltin electrodes is their functional lifetime, which is limited by
172 degradation of the response within days. A multidentate carrier with four tin centres exhibited
173 excellent selectivity to phosphate. However the lifetime of the electrode was less than one day[55
174]Some of the ISEs based on distannyl derivatives exhibited good selectivity to phosphate. However,
175 information about their lifetime has not been reported [56]. The chemical structures of the tin based
176 ionophores are given in figure 1

177 **Here figure 1**

178

179 *2.3 Potentiometric Metal complex Sensors*

180 One of the interesting systems based on metal complexes that can recognise phosphate anions is based
181 on Zn .Two Zn(II) - dinuclear systems were studied as receptors for phosphates which were obtained
182 by using two polyamino-phenolic ligands. The affinity of the metalloreceptors towards phosphate
183 sensing was evaluated in aqueous solution in a wide range of pH ($6 < \text{pH} < 10$). One of the
184 metalloreceptors was able to selectively discriminate phosphate from pyrophosphate, and on the
185 contrary another receptor exhibited opposite selectivity. The difference in the selectivity is ascribed to
186 the different Zn(II)-Zn(II) distances between the two metal centres. The potentiometric results have
187 been substantiated by studying the interactions of phosphate with the Zn complexes through NMR
188 and fluorescence measurements of.[30] Cobalt phthalocyanine complex was used as an ionophore for
189 phosphate, which gave interesting selectivities .[57,58] ISEs with membranes containing mixed
190 ligand Ni(II) complexes (Ni[dike][diam] where dike = β -diketonate, diam = *N,N'*-di-, tri-, or tetra-
191 alkylated ethylenediamine) were selective to phosphate with response slopes of -21 mV/decade
192 [59,60]

193

194

195

196 *2.4 Phosphate ISEs based on salophens*

197 Uranyl and vanadyl salophens are used as phosphate ionophores in ion selective membranes
198 which exhibit tolerable phosphate selectivity. Their inadequate stability and short lifetime has
199 forbidden their application in direct use for environmental and clinical analysis. Moreover, they are
200 only functional under strict laboratory conditions. To circumvent these problems efforts have been
201 made to prepare terthiophene monomer appended uranysalophen, followed by polymerizing its
202 modified monomers to produce functionalized conducting polymer films (CP-ISE). The CP-ISEs
203 showed better electrochemical properties (response time, Nernstian slope and selectivity) for
204 monohydrogenphosphate over conventional ISEs incorporated with the same ionophore. Furthermore,
205 we can resort to miniaturization with the CP-ISEs since they did not require plasticized-PVC
206 membranes with internal solutions which are needed in the conventional ISEs. The CP-based
207 membrane exhibited excellent functional properties for the ion-to-electron transducers and provided
208 ion-recognition sites for the selective complexation in solid-state ISEs. However this method also was
209 not very successful due to the short life of the sensor. [61]

210 *2.5 Potentiometric sensors based on polyamines , guanidinium and ammonium receptors for*
211 *phosphate*

212 Polyamines form a special group of phosphate carriers because they have no metal center.[62] Among
213 four macrocyclic polyamines, a macrocycle with one secondary amine and two lactam groups was
214 claimed to give the highest selectivity for phosphate, giving a Nernstian response down to 10^{-6} M
215 HPO_4^{2-} . Another group of workers have used the same polyamine and have demonstrated phosphate
216 sensing in macro and microelectrodes. [63]

217 A zwitterionic bis(guanidinium) ionophore bearing an anionic closo-borane cluster which can
218 complex and selectively extract oxoanions has been investigated in polymeric membrane ion
219 selective electrodes (ISEs). By systematic variation of the concentration of the ion-exchanger sites in
220 the membrane, a reasonably good selectivity for monohydrogenphosphate was obtained. A detection
221 limit of 8.7×10^{-8} M has been reported. [64]

222 The design and synthesis of receptors containing a Cu(II) binding site with appended
223 ammonium groups and guanidinium groups , along with thermodynamic analyses of anion binding,
224 are reported. Both receptors show high affinities (10^4 M^{-1}) and selectivities for phosphate over other
225 anions in 98:2 water: methanol at biological pH. However the authors have not used these compounds
226

8

227 in ISEs. [65,66] Table 1 provides the performance of some potentiometric sensors in terms of
228 analytical parameters like detection limit, sensitivity, response time storage life, nernstian slope value
229 etc.,

230 *2.6 Innovative modifications of potentiometric analysis*

231 Established history of potentiometric sensors, accompanied with their simple instrumentation
232 requirement and low production costs, make them attractive analytical tools suitable for a wide variety
233 of applications. However analysis of very small concentration will result in insignificant changes in
234 potentials which make the determinations prone to errors. A 10% activity change of a monovalent
235 cation at 25 °C leads to a mere 2.4 mV change in the emf as predicted by Nernst equation. When the
236 activity is doubled emf changes by 17.8 mV. Temperature changes and improper reference electrode
237 themselves will give rise to an error of similar magnitude. Hence proper control of temperature,
238 frequent recalibrations and use of highly reliable reference electrodes are mandatory for
239 potentiometric analysis. These problems remain as stumbling blocks for the implementation of
240 potentiometric sensors for applications as implantable electrodes and for remote sensing using
241 wireless sensor networks. Hence researchers have come out with some modifications of the
242 potentiometric analysis to overcome such obstacles. Some of the innovative modifications of
243 potentiometric analysis attempted are high-amplitude sensing, backside calibration potentiometry,
244 constant current coulometry and coulometric ion transfer. [67] These modifications are expected to
245 extend the applications of potentiometric sensors to remote sensing and for the fabrication of
246 implantable electrodes that can function for prolonged periods. [67]

247 **3. Voltammetric Detection of phosphate**

248 *3.1 Voltammetric methods based on supramolecular recognition of phosphate*

249 Designing of a good anion receptor requires selection of a proper signal unit and designing of
250 an effective binding site in case of sensing of anions by voltammetry. Potentiometric determination
251 does not require a signalling unit as it depends on the distribution of anions in the membrane
252 containing the recognition molecule with the binding sites and the analyte solution and the resulting
253 potential changes (nernstian) at the interface. The factors to be considered while designing a selective
254 receptor for anions are geometry, basicity of the anion and the nature of the solvent medium where

255 sensing has to be carried out. Complementarity between the receptor and anion is highly crucial in
256 determining selectivities. A useful way of grouping anion receptors is to consider the types of
257 noncovalent interaction used to complex the anionic guest. These include electrostatic interactions,
258 hydrogen bonding, hydrophobicity, coordination to a metal ion, and combinations of these interactions
259 working together. Electrochemical molecular recognition is an expanding research area at the
260 interface of electrochemistry and supramolecular chemistry.[29, 30] Schematic diagrams of this
261 approach are shown in Figure 2. A variety of organic, organometallic, and inorganic redox active
262 centres (signal units) have been incorporated into various molecular recognition frameworks
263 containing the binding sites and have been shown to electrochemically detect anions. Electrochemical
264 interrogation of phosphate anions by techniques such as cyclic voltammetry (CV) has been widely
265 used in anion recognition for its own advantages like convenience in its operation, low cost and small
266 sample volumes.

267 **Here Figure 2**

268 *3.1.a Metallocene Receptor systems*

269 *Cobaltocene based receptors*

270 The first redox-active class of anion receptors, based on the cobaltocenium moiety was
271 reported by Beer and Keefe in 1989.[68] Since then, a plethora of acyclic, macrocyclic, and calixarene
272 receptors containing cobaltocenium(Cp_2Co) have been prepared. Cyclic voltammetric experiments
273 demonstrated that all these receptors could electrochemically sense anions. The addition of anions to
274 solutions of the receptors in acetonitrile resulted in significant shift of the reversible Cp_2Co^+/Cp_2Co
275 redox couple towards lower potentials. The complexed anionic guest effectively stabilizes the
276 positively charged cobalt centre making it more difficult to reduce. For example, complexation of
277 chloride ions by amide functionalised cobaltocenium receptors induced shifts between 30 mV and 85
278 mV, whereas larger magnitudes of 200 mV and 240 mV were observed for the complexation of
279 dihydrogenphosphate towards lower potentials. [69, 70] The anion-coordination properties of the
280 cobaltocenium bridged calix[4]arene receptors are dependent upon the degree of preorganization of
281 the upper rim. Recognition can be tuned in favour of phosphate anions by exchanging the positions of

282 the tosyl substituent on the lower rim of the calix[4]arene which had a dramatic influence on the
283 anion coordination properties of the upper rim.[71-73]

284 ***Ferrocene based receptors***

285 In the electrochemical anion recognition area, especially with respect to phosphate anions,
286 ferrocene group has proved to be a very good signal unit because of its stable electrochemical
287 properties and its ease of detection by methods such as cyclic voltammetry (CV). Besides, the
288 ferrocene group can modulate the binding event through alternating between its two redox states. As
289 for binding site, amide, [74] urea, [75] and the hydroxyl group [76] that can form hydrogen bonds
290 with anions are commonly used. Quarternized nitrogen [77] and positively charged pyridine [78] are
291 also chosen as binding sites on the basis of the electrostatic interaction. Others that can provide shape
292 complementation [79] are also considered. Ferrocenyl esters such as glycidyl ester of ferrocene
293 carboxylate (GEFC) and 1,3-diferrocenecarboxylic acid diacylglycerol (DFCDG), 1,1'-*N,N'*-
294 ferrocenoyl bisamino acid methyl esters and bisferrocenyl-substituted urea and thiourea and
295 trisferrocenyl-substituted guanidine derivatives were also evaluated for sensing anions.[80-82]
296 However most of the receptors respond not only to phosphate but also to anions like bisulphate,
297 fluoride anions etc., Hence sensitivity and selectivity are still subjects of further investigations. It is
298 now well accepted that one single interaction in the receptor molecule such as hydrogen bond or
299 electrostatic interaction or shape complementation may not be enough to improve the selectivity and
300 sensitivity for phosphate, especially in an aqueous environment.[83] Thus, combination of several
301 interactions is being considered. It is believed that multiple binding sites involving several different
302 binding groups such as those that can form hydrogen bond or that can provide electrostatic
303 interactions, will improve the sensitivity and selectivity of anion receptors. A new ferrocenyl anion
304 receptor with specially designed multiple binding sites viz., amide and positively charged nitrogen
305 (*N,N,N,N*-(dimethyl, ethyl, ferrocenecarboxylic amidodimethylene) ammonium fluoborate) was
306 synthesized by Tan et al.[84] Compared to its counterpart with just a single binding site of amide, this
307 compound with multiple binding sites showed higher sensitivity to H_2PO_4^- and thus proved the
308 enhancement effect of the multiple binding sites.[84] Ferrocene substituted calix(4)pyrroles have been

309 synthesized and investigated using acetonitrile: DMSO mixture (9:1) by cyclic voltammetry and
310 square wave voltammetry and it was found to bind fluoride, chloride and dihydrogenphosphate
311 anions. [85] A neutral redox-active receptor (ferrocene functionalized calix[4]pyrrole) was used as an
312 active component in carbon paste electrodes as ion-selective electrodes (ISEs), for the detection of
313 anions in aqueous solution. Measurements with carbon paste electrodes were conducted using
314 Osteryoung square-wave voltammetry. Amongst the anions studied, dihydrogenphosphate and
315 fluoride caused the strongest decrease of peak current (approximately 25%), followed by bromide and
316 chloride.[86] The electropolymerization of a simple monoamidoferrocene derivative containing a
317 pyrrole group is a straightforward way to synthesize redox polymer films that can sense H_2PO_4^- ,
318 ATP^{2-} and HSO_4^- , with excellent selectivity for the former anions.[87] Platinum and gold
319 microelectrode arrays (MEAs), fabricated on silicon substrates with different geometric
320 characteristics, were surface-modified by the potentiostatic electropolymerization of the pyrrole-
321 ferrocene derivative, in the case of the platinum MEAs, and by the chemisorption of the thiol-
322 functionalized ferrocene, in the case of the gold MEAs. The modified MEAs were investigated for the
323 detection of the dihydrogenphosphate mono-anion in nonaqueous media via differential pulse
324 voltammetry. This was based on electrostatic interactions and/ or hydrogen bonding between the
325 target anion and the amide-ferrocene or ammonium-ferrocene functionalized electrode surfaces. A
326 decrease in the ferrocene (Fc) oxidation peak current with a concomitant increase in the peak current
327 of a new peak at lower potentials was observed when the concentration of the dihydrogenphosphate
328 was increased. This method exhibited very good selectivity for H_2PO_4^- anions compared to ATP,
329 HSO_4^- and NO_3^- ions and the analysis was performed in nonaqueous solution using differential pulse
330 voltammetry.[88] Pentamethyl amidoferrocene dendrimers and silane based ferrocene dendrimers (in
331 solution and in the modified phase) have also been evaluated for the recognition of anions like
332 phosphate, ATP, HSO_4^- etc. [89,90] . Amide substituted tetrathiafulvalene derivatives are used with
333 some success for the selective determination of H_2PO_4^- over other anions in nonaqueous
334 medium.[91] The main disadvantage of all these systems is the lack of selectivity in the majority of
335 the cases and the studies are confined only to aprotic media with one or two exceptions.[86,87,92]

336 The reference [87] discusses selective detection of phosphate in aqueous environment whereas the
337 references [86] and [92] discuss selective sensing of phosphate in non aqueous conditions.

338 *3.1.b Anion Complexation through Second-Sphere Coordination*

339 ***Transition-Metal Bipyridyl Based Receptors***

340 The use of a second coordination sphere of metal complexes as a basis of anion recognition is
341 another method for the development of hydrogen-bond based inorganic anion receptor. The redox-
342 active and photoactive ruthenium(II) bipyridyl moiety, in combination with secondary amide groups,
343 incorporated into acyclic, macrocyclic, and lower-rim calix[4]arene structural frameworks are shown
344 to produce a new class of anion receptors capable of optical and electrochemical sensing. [93].
345 Single-crystal X-ray structures of the H_2PO_4^- complex of the Ru(II) bipyridyl compound in
346 combination with secondary amide groups incorporated into calixarene frame work highlight the
347 importance of hydrogen bonding to the overall second sphere anion complexation process. Three
348 hydrogen bonds (two amide and one calix[4]arene hydroxyl) stabilize the H_2PO_4^- anion. The
349 ruthenium ion is dipositive, and hence, electrostatic interactions are particularly favorable. The
350 macrocyclic receptors form highly selective and thermodynamically stable complexes with H_2PO_4^- .
351 Electrochemical anion recognition experiments showed substantial anion-induced cathodic
352 perturbation of the phosphate complex in agreement with stability constant values of $28\,000\text{ M}^{-1}$ for
353 H_2PO_4^- in DMSO, capable of selectively sensing H_2PO_4^- in the presence of 10-fold excess amounts of
354 HSO_4^- and Cl^- .

355 *3.1.c Heteroditopic sensing*

356 The design of heteroditopic ligands that contain two quite different binding sites for the
357 simultaneous complexation of cationic and anionic guest species is an emerging field of
358 supramolecular chemistry. These multisite ligands are able to bind a single heteroditopic guest or
359 simultaneously bind two non-identical guests. The invention of convergent heteroditopic hosts is a
360 challenging problem in molecular design because the binding sites have to be incorporated into a
361 suitably preorganized scaffold that holds them in close proximity, but not so close that sites interact.
362 Ferrocene-based ionophores substituted with crown ethers or polyaza-macrocycles exhibit interesting

363 electrochemical cation recognition effects because the complexing ability of the ligand can be
364 switched on and off by varying the applied electrochemical potential. Owing to the relatively strong
365 hydrogen bonding ability of the urea group, a number of molecules possessing the urea motif have
366 been designed as neutral receptors for various anions. By combining the redox activity of the
367 ferrocene moiety with the anion binding ability of the urea group and a crown ether moiety as an
368 alkaline metal- binding site, a new heteroditopic ferrocene based ligand capable of the simultaneous
369 binding of anions and cations can be designed. A heteroditopic ligand containing urea and a crown
370 ether group synthesized by F.Ot' on et al. was studied by cyclic voltammetry (CV) in dichloromethane
371 containing 0.1 M TBAClO₄ as supporting electrolyte. The compound exhibited a reversible one-
372 electron oxidation process corresponding to ferrocenium-ferrocene (Fc⁺/Fc) couple. Electrochemical
373 anion and cation sensing experiments were carried out by differential pulse voltammetry (DPV). On
374 stepwise addition of 1.5 equivalents of F⁻ (as its TBA⁺ salt) led to a modest cathodic shift of -52 mV.
375 However, upon addition of 2 equivalents of H₂PO₄⁻ a very large shift of 190mV occurred in the
376 negative direction, reflecting a strong binding of the guest upon oxidation of the ferrocene unit.
377 Maximum perturbation of the differential pulse voltammetry (DPV) output was obtained with 2
378 equivalents of added H₂PO₄⁻ anion. Remarkably, the presence of Cl⁻, Br⁻, HSO₄⁻ and NO₃⁻ anions
379 had no effect on the DPV, even when present in large excess.[94] Guo et al. synthesised a ferrocene-
380 based 1,3-alternate thiacalix[4]arene ditopic receptor that contained four identical polyether arms
381 terminated with the ferrocene amide moieties. Cyclic voltammetric studies conducted in a nonaqueous
382 medium containing 1:1 dichloromethane and acetonitrile have revealed that this redox-active receptor
383 can be used as an electrochemical sensor to recognize both europium (Eu³⁺) and dihydrogenphosphate
384 H₂PO₄⁻ ions with a high selectivity. [92] P.D.Beer et al. demonstrated using water soluble pH
385 dependent polyazaferrocene macrocyclic ligands that the pH dependent electrochemical recognition
386 of transition metal cations and phosphate anions in the aqueous environment. At low pH, the
387 compound exists in the protonated form and can be used to determine biologically important anions
388 like phosphate and ATP in the aqueous environment. At high pH they exhibit recognition properties
389 towards cations especially with respect to Cu²⁺ ions. [95]

390

391 *3.2 Voltammetry at the interface between two immiscible electrolyte interfaces (ITIES)*

392

393 Many calixarene compounds are known to be anion-selective towards halides (Cl^- , Br^- , I^-)
394 [96-98]. Of late the use of modified calix[4]arene and calix[6]arene molecules have effectively been
395 used for phosphate sensing in PVC-membrane ion selective electrodes (ISEs) [99,100]. The urea
396 functionalised calixarene was demonstrated to exhibit selectivity towards phosphate compared to
397 common anions like sulphate and chloride. However anions like nitrate and perchlorate interfered in
398 the detection. [36] The disadvantages associated with interference using ISEs may be overcome by the
399 introduction of a supplementary measurement dimension. The ions that cannot be distinguished under
400 equilibrium potentiometry conditions can be analysed with the help of voltammetric or amperometric
401 techniques at the liquid-liquid interface which allows the separation of co-transferring ions on the
402 potential axis. This leads to an improvement in the performance of the sensing process. The
403 interaction of a urea-functionalized calix[4]arene ionophore and phosphate was investigated by
404 voltammetric ion transfer at the interface between two immiscible electrolyte solutions (ITIES).
405 Voltammetry at the ITIES established that the ionophore-facilitated transfer of
406 monohydrogenphosphate occurred in preference to dihydrogenphosphate transfer. The results are
407 comparable with previously reported data on the potentiometric evaluation of this calixarene as an
408 ionophore in PVC-membrane electrodes.[101] The data provide the foundation for the development of
409 amperometric monohydrogenphosphate sensors based on the ion-transfer principle

410

411 *3.3 Thermal modulation (TM) voltammetry*

412

413 Thermal modulation (TM) voltammetry involves the determination of changes in the
414 voltammetric signals ($\Delta I/\Delta V$), when the electrode/electrolyte surface is periodically heated using laser
415 sources. The thermal modulation of the interface changes the standard entropy of the electrode
416 reaction. The temperature coefficient of the standard potential, ($\partial E^\circ/\partial T$), for the electrode reaction of

15

417 Ox + ne ⇌ Red is equal to the product of the number of electrons, the Faraday constant and the
418 standard entropy change, $-nF\Delta S^\circ$. Subsequently, when the temperature is increased from T to $T + \Delta T$,
419 and if the electrode reaction has a positive value of the standard entropy change, the standard potential
420 shifts to a negative direction, and *vice versa*. Since the limiting current increases with the diffusion
421 coefficient, and hence with the temperature, the limiting current at $T + \Delta T$ is always greater than that
422 at T . Such temperature effects on thermodynamics and diffusion lead to a minor difference between
423 two voltammograms at T and $T + \Delta T$, where ΔT is much less than T . TM voltammetry is a sensitive
424 enough to detect such small differences. The measurement involves a periodic heating and lock-in
425 detection system in addition to the conventional voltammetric instruments. TM voltammetry has been
426 explored for the detection of phosphate in natural water samples by using a He-Cd dual laser as a
427 heating source and a graphite-reinforced carbon (GRC) electrode. The heteropoly ion, i.e., 12-
428 molybdophosphate ion ($[\text{PMo(VI)}_{12}\text{O}_{40}]^{3-}$), was formed through a reaction between phosphate and
429 molybdate ions in an acidic solution, and its electroreduction was investigated in a flow electrolytic
430 cell by TM voltammetry. [102] Measured TM voltammograms showed two peaks corresponding to
431 two successive two-electron reductions of the 12-molybdophosphate ion, and the peak intensities were
432 proportional to the concentration of the phosphate ion. Because of the strong adsorption of 12-
433 molybdophosphate ion onto the graphite reinforced carbon electrode, a detection limit as low as
434 $0.8 \times 10^{-9} \text{ mol L}^{-1}$ was achieved. The determination of phosphate ion in river water was carried out by
435 TM Voltammetry The results obtained were similar to those obtained by the spectrophotometric
436 molybdenum blue method. These results prove the significance of TM voltammetry as an
437 electroanalytical method for the determination of phosphate.[102]

438

439 **4. Amperometric detection of phosphate**

440

441 *4.1 Amperometric detection via electrochemical reduction of phosphomolybdates*

442 The popular analytical method for the determination of phosphate involves treatment of the
443 sample with an acidic molybdate solution to convert the phosphate into the Keggin anions ($\text{PMo}_{12}\text{O}_{40}$)

16

444 ³⁻) and subsequent chemical reduction leading to mixed molybdenum oxidation state.[103] These ions
445 are intensely blue colored which allows spectrophotometric determination of trace level phosphate in
446 an analyte. As a routine analytical method, this chemistry is carried out in an automated continuous
447 flow assembly. The chemistry of these Keggin anions is complex and the rate of formation, stability
448 and ratio of isomers depend strongly on the solution conditions, including pH, and the method suffers
449 from interferences from silicate, arsenate etc. Sometimes addition of organic solvents is required for
450 the selective extraction of the desired analyte. Even the order of addition of reagents affects the results
451 of the experiment and therefore the method is not entirely and universally satisfactory. To avoid the
452 complications arising from the use of chemical reducing agents, the keggin anions are reduced
453 electrochemically followed by spectrophotometric analysis. This technique has successfully been
454 employed for the determination of orthophosphates in beverages, waste waters and urine samples
455 [103]. FIA (flow injection analysis) and spectrophotometric techniques, commonly used for the
456 measurement of phosphate in the laboratory, are not suitable for on-site testing and monitoring. The
457 use of direct electroreduction techniques offer portability and excellent sensitivity which make them
458 very attractive for on site monitoring of phosphate. Furthermore the interference from ions like
459 silicate and arsenate can be completely avoided as reductions of their respective species occur at
460 different potentials. The potential dependent selectivity combined with the portability and prospect of
461 miniaturisation make the electrochemical determination of phosphate using molybdophosphate highly
462 versatile. Several reports on the electrochemical determination of phosphate using
463 phosphomolybdates are available in the literature. [5,104-110] Amperometric detection in flow
464 injection analysis is widely reported in the literature. [5,107-110] One of the papers describes
465 formation of phosphomolybdate complex in presence of nitric acid, ammonium molybdate and
466 phosphate and its subsequent reduction at a carbon paste electrode, polarised at +0.3V (versus
467 Ag/AgCl) .[5] The major characteristics of the method were simplicity of the equipment, limited
468 consumption of reagents and low limit of detection (0.3×10^{-6} mol L⁻¹) with a linear range between 1
469 and 20×10^{-6} mol L⁻¹. The interference of silicate was completely eliminated by using appropriate
470 concentrations of nitric acid and ammonium molybdate. This method was successfully applied to

471 orthophosphate analysis in cyanobacterial biofilms collected from Roman catacombs.[5] The potential
 472 dependent selective determination of silicates and phosphates was also evaluated by carrying out the
 473 voltammetry of the molybdosilicate and molybdophosphate complexes, formed by the addition of
 474 hexafluorosilicate and phosphate to an acidic sodium molybdate solution, at gold microdisk
 475 electrodes.[109] It is shown that the reaction conditions influence both the kinetics of formation of the
 476 complexes and their voltammetry. It is possible to find the conditions where the steady state
 477 amperometric response of the Au microdisk electrodes allows a rapid and convenient method for the
 478 determination of silicate and phosphate at concentrations in the range $1 - 1000 \times 10^{-6} \text{ mol L}^{-1}$. [111].
 479 However from the perspective of researchers involved in remote sensing, it has been reported that the
 480 colorimetric analysis of phosphate based on ammonium molybdate meets the stringent analytical
 481 requirements needed for remote sensing. The power requirement for the colorimetric detector is also
 482 suitable for remote sensing. Similar ruggedness can also be developed in the case of electrochemical
 483 sensing of phosphate using ammonium molybdate [112]. The reaction occurring between molybdate
 484 ions and phosphate ions resulting in blue colour is as follows:



488
489
490

491 A perovskite-type oxide-based electrode showed good properties of amperometric sensing to
 492 hydrogen-phosphate ion. The carbon electrode loaded with $\text{La}_{0.9}\text{Ce}_{0.1}\text{CoO}_3$ showed remarkable
 493 selectivity to HPO_4^{2-} among the examined anions of F^- , Cl^- , Br^- , SCN^- , NO_3^- , SO_4^{2-} , CO_3^{2-} and ClO_4^- ,
 494 although it received serious interference from I^- . The LaCoO_3 thin film sensor device responded to
 495 HPO_4^{2-} at concentrations between 1.0×10^{-6} and $1.0 \times 10^{-1} \text{ mol L}^{-1}$. [113]

496

497 4.2 Indirect determination of phosphate

498 A highly selective enzymeless approach using a $\text{Ni}(\text{OH})_2/\text{NiO}(\text{OH})$ modified barrel plated
 499 nickel electrode (Ni-BPE) in alkaline media for the determination of phosphate (PO_4^{3-}) by flow
 500 injection analysis (FIA) has been reported recently.[114] The presence of $\text{Ni}(\text{OH})_2/\text{NiOOH}$

501 activates the adsorption of phosphate at the electrode surface which inhibits the current
502 corresponding to the electrocatalytic oxidation of glucose in 0.1 M NaOH solution. Under the
503 optimized conditions of flow rate (300 $\mu\text{L}/\text{min}$), detection potential (0.55 V vs Ag/AgCl) and with 25
504 $\times 10^{-6}$ mol L⁻¹ glucose in 0.1 M NaOH as carrier solution, the calibration curve showed a linear range
505 up to 1×10^{-3} molL⁻¹. Probable interference from coexisting ions was also examined. The results
506 confirmed that the sensor could be used for the determination of phosphate in the presence of nitrate,
507 chloride, sulfate, acetate, oxalate, carbonate. It could also be used in the presence of other anionic
508 species of toxicological and environmental interest, such as chlorate, chromate, and arsenate ions. The
509 electrode could be efficiently regenerated without further treatment under the hydrodynamic
510 condition. For eight continuous injections of 40×10^{-6} mol L⁻¹ PO₄³⁻, a relative standard deviation of
511 0.28% was obtained, indicating good reproducibility of the proposed method. A detection limit of 0.3
512 $\times 10^{-6}$ mol L⁻¹ was achieved by this method. A schematic diagram of the sensing mechanism is given
513 in figure 3.

514 **Here figure 3**

515

516 *4.3 Amperometry coupled with ion chromatography*

517 An amperometric sensor intended especially for non-electroactive ions, functioning under
518 flow injection mode, was applied as a novel detector in suppressed ion chromatography. It consists of
519 a carbon paste electrode modified with either a polycationic or a polyanionic polymer holding a
520 suitable charge transfer mediator ([Fe(CN)₆]³⁻ or Cu²⁺), functioning in an indirect amperometric mode.
521 The detection mechanism involves ion exchange between the non-redox ionic analyte and the
522 electroactive mediator, in the polymer particles positioned at the electrode surface, followed by the
523 electrochemical transformation of the mediator species leached out of the polymer at the electrode /
524 solution interface. The estimation was accomplished in the absence of added supporting electrolyte.
525 Optimisation was performed to get the highest faradic signals, by varying a range of experimental
526 parameters (i.e. applied potential, composition of the electrode). These systems were then successfully
527 applied to the analysis of mixtures of cations (Li⁺, Na⁺, K⁺, NH₄⁺, Ca²⁺, Mg²⁺) and anions (F⁻, Cl⁻,

528 NO_2^- , NO_3^- , SO_4^{2-} , PO_4^{3-} following chromatographic separation. Good operational stability was
529 observed, with typically less than 5% signal loss for 50 consecutive measurements.[115]

530

531 **5. Sensing of phosphate using self- assembled monolayers**

532 Self-assembled monolayers (SAMs) of thiol-derivatized molecules on gold substrates have
533 recently received substantial consideration in connection with their potential applications for
534 sophisticated designs of molecular-based electronics, chemical sensors, and nanopatterning. Reports
535 on SAM based platforms for phosphate sensing is available in the literature. [116,117] In one of the
536 reports, ferrocene based thiols, appended with binding site for oxoanions (most often amide groups or
537 trialkyl ammonium groups.) are self-assembled on gold electrodes and the sensing of phosphate
538 anions are evaluated in organic and inorganic media using voltammetric shifts produced for the
539 ferrocene/ferrocenium couple as a result of binding the anions. [116,117] However this method
540 responded to several anions and is not selective.

541 In another case the blocking of electron transfer properties for ferrocyanide electron transfer
542 across a SAM modified electrode containing a binding site for anions (porphyrin or Zirconium (IV)
543 ions) was evaluated for the sensing of phosphate anions. This approach was found to be selective for
544 phosphate over many other anions. [118,119]

545 A SAM based platform derived from mercaptopropionic acid was further functionalised with
546 Zr(IV) ions and was found to be an effective modified electrode for the sensing of phosphate anions.
547 The method and applicability of the sensor were successfully tested by detection of phosphate in
548 blood serum after deproteinization of the sample without interference from the sample matrix. [119]

549 A schematic diagram of the sensing mechanism is given in figure 4.

550 **Here figure 4**

551 **6. Unconventional methods for detection of phosphate**

552 The detection of phosphate ions by using certain unconventional methods has also been
553 reported in the literature. Some of these are, a) following the intensity of electrochemiluminescence
554 changes of the complex formed between molybdophosphates and a luminescent dye. [34] For

555 example, the electrochemiluminescence (ECL) method was carried out with a hydrophobic ion
556 dissociated complex formed between molybdophosphoric heteropolyacid and protonated
557 butylrhodamine B (BRhB). The complex was selectively extracted into the bulk of paraffin oil based
558 carbon paste electrode. The (ECL) was followed at +1.3V in alkaline medium. Under the optimum
559 experimental conditions, the ECL intensity was linear with the concentration of phosphate (as
560 phosphorus) in the range of 6.4×10^{-9} to 1.0×10^{-7} mol L⁻¹. The detection limit was 2.5×10^{-12} mol L⁻¹.
561 The proposed method has been applied successfully to the analysis of phosphate in the water samples.
562 [34] b) following the reduction in transmittances of cobalt oxide thin film electrodes induced by the
563 adsorption of phosphate at fixed potential [120] c) following the mass changes associated with the
564 inclusion of phosphate anions in conducting polymer films like PEDOT(polyethylenedioxythiophene)
565 using a quartz crystal microbalance, QCM. [121] d) a microfluidics-based ion-channel sensing system
566 for nonelectroactive anions under negative separation electric field that relies on amperometric
567 response of the oxidation of a carbon fiber electrode.[122]

568

569 **7. Bioelectroanalytical detection of phosphate**

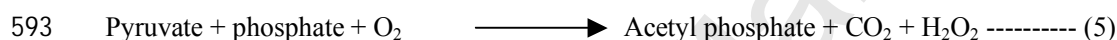
570 Biosensors for the determination of phosphate are normally based on mono- or multi-
571 enzymatic reactions where phosphate acts as inhibitor or substrate. The enzyme-based amperometric
572 biosensors are highly advantageous due to the high selectivity of the biorecognition element and the
573 sensitivity of the electrochemical signal transduction. Despite the promise of high selectivity,
574 phosphate selective enzymes are not readily amenable to direct electrochemical interrogation. One of
575 the enzymes often employed for the sensing of phosphate is pyruvate oxidase (POD). The reaction
576 involves the conversion of pyruvate to acetyl phosphate in the presence of oxygen to yield hydrogen
577 peroxide. The concentration of phosphate is inferred from the amperometric measurement of either
578 oxygen depletion or the increase in concentration of hydrogen peroxide. [4, 20,123-125] These
579 detection modes have a number of limitations. Measurement of a stable and clear-cut signal for the
580 reduction of oxygen is often difficult and will be tricky in situations where the concentration of
581 oxygen is very low. The oxidation of peroxide is hampered by poor electrode kinetics at conventional

582 electrode materials and requires a large over potential for oxidation to give rise to a quantifiable
 583 current signal and hence prone to interferences from oxidisable impurities in the solution. In certain
 584 cases where oxygen concentration is a limiting factor, mediators are employed for the detection of
 585 phosphate. Osmium bipyridyl units functionalised pyrrole monomer, copolymerised electrochemically
 586 with thiophene was used as a mediator for pyruvate oxidase electron transfer and the detection of
 587 phosphate was demonstrated successfully using this mediator. [125] The sensitivity of detection
 588 achieved in this case is 0.2 A cm^{-2} and detection range is between 0.02 mol L^{-1} and 5 mol L^{-1} . The
 589 POD catalysed enzymatic reaction with phosphate as the substrate is as follows:

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592



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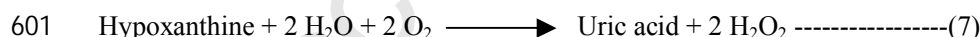
596 Nucleoside phosphorylase has been used [126,127] for the sensing of inorganic
 597 orthophosphate based on the following reactions

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598



600



602

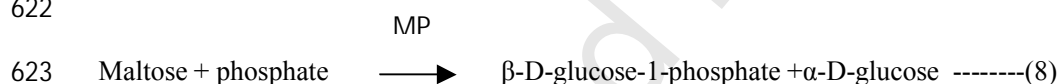
603 The concentration of phosphate was followed by studying the consumption of oxygen caused by the
 604 reaction with hypoxanthine which is generated during the enzymatic reaction between inosine and
 605 orthophosphate. The enzyme was immobilized on a membrane prepared from cellulose triacetate and
 606 was fixed on top of a Clark-type oxygen electrode. A detection limit of $10^{-4} \text{ mol L}^{-1}$ phosphate was
 607 achieved by this method. Later d'Urso and Coulet [128] and Haemmerli et al.[129] were able to
 608 increase the sensitivity of this method by using a hydrogen peroxide transducer instead of using an

609 oxygen electrode and a detection limit of 10^{-7} mol L⁻¹ was confirmed. The ability to make use of the
 610 uric acid signal provides an important operational advantage. Urate is endogenous to physiological
 611 systems and hence would prove to be a substantial interferent in actual analysis of clinical samples
 612 [23]. In the context of environmental analysis, it is likely that only certain samples would be expected
 613 to contain the purine and hence the direct oxidation of the base at the electrode can be assumed to be
 614 derived solely from the enzymatic sensor assembly. The advantage of exploiting this label rather than
 615 peroxide lies in the relatively low oxidation potential of the purine ($\sim+0.2$ to $+0.5$ V). The oxidation
 616 of peroxide is associated with poor electrode kinetics and large over potentials ($\sim+0.8$ to $+1$ V vs.
 617 Ag/AgCl) at conventional electrode substrates.

618

619 An enzymatic sensor based on four different enzymes for phosphate detection was reported
 620 using maltose phosphorylase (MP), acid phosphatase (AP), glucose oxidase (GOD) and mutarotase
 621 (MR) with the following reaction sequence [130].

622



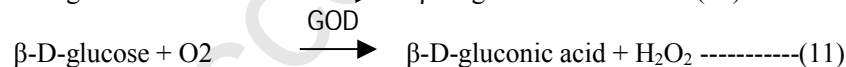
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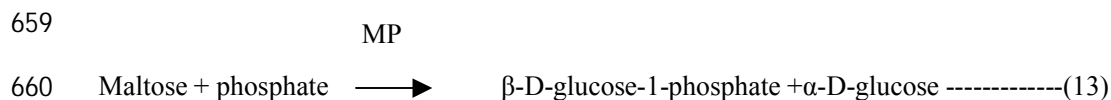
631

632 The combination of the former two enzymes generates two glucose molecules per reaction cycle and
 633 recycles one molecule of phosphate, and the oxidation of glucose is catalyzed by the glucose oxidase
 634 enzyme after its mutarotation. The formation of hydrogen peroxide during the enzymatic reaction can
 635 be monitored electrochemically at a platinum electrode. Based on this study, both tri- and bi-

23

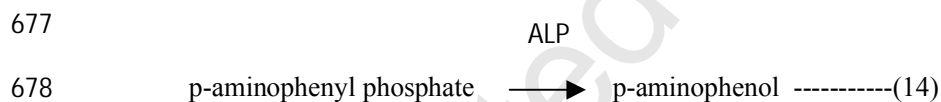
636 enzymatic sensors for phosphate detection have been described. Mousty et al. [19] used a simple
 637 method to fabricate an amperometric phosphate biosensor containing MP, MR, and GOD with a linear
 638 range of $1-50 \times 10^{-6} \text{ mol L}^{-1}$. Huwiel et al. [131] successfully applied MP and GOD as the bioelements
 639 of a simple bi-enzymatic sensor, in which the first enzyme consumed phosphate as a cosubstrate and
 640 yielded a product that was a substrate for the second enzyme. However, more enzymes involved in the
 641 sensor system lead to more non-specific response due to the presence of substrates for the next
 642 enzymes. Furthermore, the instability of each enzyme caused fluctuations in the sensor performances.
 643 For example, in the phosphate detection system consisting of nucleoside phosphorylase and xanthine
 644 oxidase, the degradation of inosine often restricted the dependability of phosphate analysis [132]. For
 645 this bi-enzyme system, no linear range was found even though it sensed phosphate [132,133]. Further
 646 immobilisation of three to four enzymes in spatially separated planes is a challenging task. Utmost
 647 care should be taken to keep the enzymes in the active state in the immobilised conditions in the
 648 presence of other enzymes. When more enzymes are present, the system becomes complicated and the
 649 results will be difficult to understand and troubleshooting will be hardly practicable. Fouling at the
 650 electrode surface will be another issue to be addressed in this case. Hence, multi-enzyme systems are
 651 rather complicated and expensive.

652
 653 Based on a monoenzymatic reaction, Zhang et al. [134] developed a conductometric biosensor which
 654 measures the conductance changes associated with the following reaction on the addition of
 655 phosphate. The detection limit achieved was $1 \times 10^{-6} \text{ mol L}^{-1}$. No interference from other anionic
 656 species was detected. The conductometric biosensor exhibited a long-term storage and operational
 657 stability as well as a good thermal stability. Measurements in the real water samples were satisfactory.
 658 The enzymatic reaction is as follows;



662 Another enzyme that is often used for the determination of phosphate esters is the enzyme alkaline

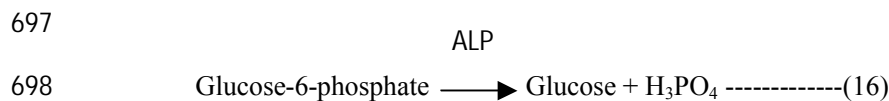
663 phosphatase (ALP) and it is mainly used for the determination of phosphate esters or for the
 664 determination of the enzyme activity. Hence the direct determination of inorganic phosphate is not
 665 possible with this enzyme. The adaptation of the methodology for the detection of orthophosphate
 666 relies upon the inhibitory action of the latter on the hydrolysis of the ester substrate. The sample is
 667 assayed using known concentrations of enzyme/substrate with the decrease of the signal from that
 668 expected in the absence of added phosphate being inversely related to the concentration of the latter.
 669 Amino phenol phosphates are generally used as the substrate for alkaline phosphatase enzyme.
 670 However the aminophenol that is generated during the enzyme hydrolysis gets oxidised only at high
 671 overpotential and this method suffers from interferences. Second issue that arises is that the
 672 electrochemical oxidation leads to polymeric deposits on the electrode. These tend to block the
 673 electrode reducing sensitivity and compromising the reproducibility of the technique. Hence
 674 phosphate esters containing ferrocene or aminophenyl derivatives that can be oxidised at low
 675 overpotentials were synthesized and explored for phosphate detection. The alkaline phosphatase
 676 catalysed enzymatic reaction using p-aminophenyl phosphate as the substrate is as follows:



681

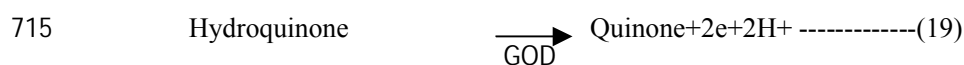
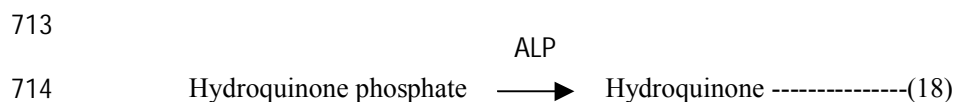
682 The majority of systems using ALP make use of a combination of enzymes in order to
 683 produce an electrochemical label with facile electron transfer kinetics. Such systems rely upon the
 684 synergistic interaction of the multi-enzyme assembly to yield a product (typically peroxide) that is
 685 more amenable to electrochemical detection than the labelled esters. A typical bienzyme system
 686 reported in the literature involved ALP/Glucose Oxidase (GOD) assemblies with glucose-6-phosphate
 687 as the key substrate in the reaction and the electrochemical label here is the oxygen that is consumed
 688 during the oxidation of glucose by glucose oxidase. Increased phosphate concentrations inhibit the
 689 production of glucose and hence the consumption of oxygen is decreased [135-137] as is the yield of

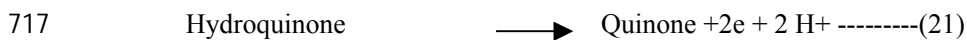
690 peroxide [138]. The multi enzyme assembly thus constructed was found to work satisfactorily within
 691 a range of environmental matrices like fresh and sea water samples. Interference from heavy metal
 692 ions (mercuric, cupric and zinc) can occur, but these are not likely to appear in any appreciable
 693 concentration in natural samples. The limit of detection for phosphate using the ALP/GOX
 694 combination was typically 0.4ppm (4 μ M) and is comparable to those obtained using the molybdate
 695 systems. The alkaline phosphatase catalysed enzymatic reaction with glucose-6-phosphate as the
 696 substrate is as follows:



701

702 Improvements in detection limit can be achieved through the catalytic cycling of the
 703 hydrolysed label. Hydroquinone monophosphate was used as the substrate for alkaline phosphatase.
 704 The hydrolysis product, hydroquinone, is capable of fast redox interconversion at the electrode
 705 surface. When ALP is combined with the glucose oxidase enzyme, in presence of an excess of
 706 glucose (to keep the FAD groups of the enzyme in the reduced state), the electrochemical oxidation
 707 product (benzoquinone) of the hydrolysis product hydroquinone, chemically reacts with the glucose
 708 oxidase and gets converted back to hydroquinone which can get oxidised at the electrode surface in a
 709 facile manner. A catalytic cycle builds up through which the current recorded at the electrode is
 710 effectively amplified. This method leads to subpicomolar detection of ALP. [139] It could be
 711 envisaged that the introduction of a sample containing phosphate would inhibit ALP. The reactions
 712 occurring using the bienzymatic approach are given as follows:



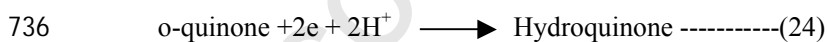
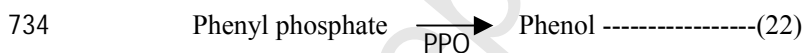


718

719 The key strength of alkaline phosphatase is that it is relatively non-specific in terms of the
 720 nature of the phosphate ester upon which it can act. This provides a significant operational advantage
 721 over some of the other enzymes in that it can be directly coupled with a wider range of secondary
 722 enzymes. Similar amplification as explained in the previous paragraph could be brought about by
 723 using phenyl phosphate as the substrate for ALP which can now be combined with the enzyme
 724 polyphenol oxidase, PPO. [140]. The hydrolysed product phenol produced by ALP reacts with PPO
 725 to yield the orthoquinone which is reduced at a cathodic potential of -0.2 V and will be free from
 726 interference effects. The current for the reduction will again be inversely related to phosphate
 727 concentration as it acts as an inhibitor for the AP catalysed ester hydrolysis. The major advantage in
 728 this instance is that oxidation of other matrix constituents can often be avoided. The PPO component
 729 therefore serves to improve both selectivity and sensitivity providing a detection limit of 0.2ppm
 730 (2×10^{-6} mol L⁻¹) for phosphate [127]. The reactions corresponding to this approach of detection is as
 731 follows:

732

733



737

738 There are a few potentiometric biosystems for phosphate.[141,142] One of the approaches has
 739 successfully harnessed the ALP enzyme-induced cleavage of phosphate. In this case the ester used as
 740 the substrate for ALP was (o-carboxyphenyl phosphate), which gave rise to salicylate as the
 741 hydrolysis product. The reaction was potentiometrically sensed with the help of salicylate membrane
 742 electrodes. The phosphate inhibited competitively the formation of salicylate to an extent inversely
 743 proportional to the concentration of added phosphate. The detection limit was 0.05×10^{-3} mol L⁻¹. The

27

744 method was successfully applied to the determination of phosphate in blood serum.[141]

745 Phosphate enzyme systems are found to be amenable for electrochemical interrogation and
746 are currently widely exploited in biomedical research. It should also be possible to transfer the
747 technology to environmental analysis as well. The multi-enzyme assemblies are however complex,
748 expensive and less stable and will be subjected to fouling by different enzymatic products. Hence in
749 order to make use of an enzymatic method of analysis advantageously, for onsite screening sites, it
750 will be preferable to make use of screen printed electrodes that can be disposed of after each analysis.
751 Table 2 provides the performance of some enzyme based sensors in terms of analytical
752 parameters like detection limit, sensitivity, response time, storage life, etc.,

753

754

755 **8. Conclusion**

756 Over the decades many analytical protocols have been developed for the sensing of
757 inorganic phosphate ions. Each method has its own limitations as mentioned in this review.
758 Potentiometric systems offer the simple requirement of instrumentation and low production costs and
759 are suitable for field based analysis, environmental monitoring, clinical analysis and remote sensing.
760 However, very small concentration changes lead to only less significant changes in the potential
761 .Hence frequent recalibrations are mandatory. Temperature should be carefully controlled to avoid
762 potential drifts arising from temperature fluctuations. The reference electrode used for the
763 measurements should be highly reliable. These are the stumbling blocks while implementing the
764 potentiometric sensors for implanting applications and remote sensing where the sensors need to be
765 kept inside the measuring locations for prolonged periods. The influence of supramolecular chemistry
766 is seen in the synthesis of a wide variety of signal compounds appended with phosphate binding
767 groups. Synthetic organic chemists are successful in preparing heteroditopic ligands that can
768 simultaneously detect an anion and a cation. Similarly a number of ferrocenyl dendrimers and silane
769 based ferrocenyl dendrimers with structural complexities have been synthesised as anion receptors.
770 Though a lot of efforts have been invested in this direction, it is generally observed that the analysis of

771 anions can only be carried out in non aqueous solvents with these supramolecular compounds and in
772 most of the cases these compounds exhibit recognition towards a number of anions. Further these
773 reports mainly showcase the organic synthetic skills of the researchers identifying supramolecules for
774 anion sensing. One of the widely used techniques that are often used for field based measurements is
775 the electrochemical sensing of phosphate using ammonium molybdate. Potential dependent selectivity
776 combined with the portability and miniaturisation capabilities make the electrochemical determination
777 of phosphate using molybdophosphate highly versatile. Literature also contains reports about some
778 non-conventional techniques that are amenable for phosphate sensing. These techniques need further
779 rigorous evaluation and input in terms of their suitability for field measurements is still required.
780 Enzymatic methods can become more popular analytical techniques for the determination of
781 phosphate when the number of enzymes participating in the detection scheme is less. Further, it is
782 preferable to use screen printed electrodes for onsite measurements and biomedical research when the
783 stability and life of the biosensor becomes questionable. However, it is possible that researchers can
784 develop genetically engineered enzymes that can exhibit biocatalytic activity for prolonged periods.
785 Advanced research is needed in the direction of enzyme based sensing for applications in biomedical
786 research for developing online monitoring systems, implantable sensors etc. In the case of
787 environmental analysis, rugged and stable sensing systems are required. Presently, the analysis of
788 phosphate, based on the electrochemical reduction of molybdophosphate and Co/Co oxide systems are
789 considered to be successful for field measurements while other methods need a lot of rigorous
790 validations. Selective sensing of phosphate is an ongoing challenge. A closer look at the problems
791 jointly by sensor chemists and anion coordination chemists is required to design ionophores for the
792 selective sensing of phosphate ions devoid of interferences from anions like nitrate and sulphate
793 which are often difficult to decouple during phosphate sensing.

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800 **References**

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Figure Captions

Figure1 Tin based ionophores reported in the literature for sensing phosphate

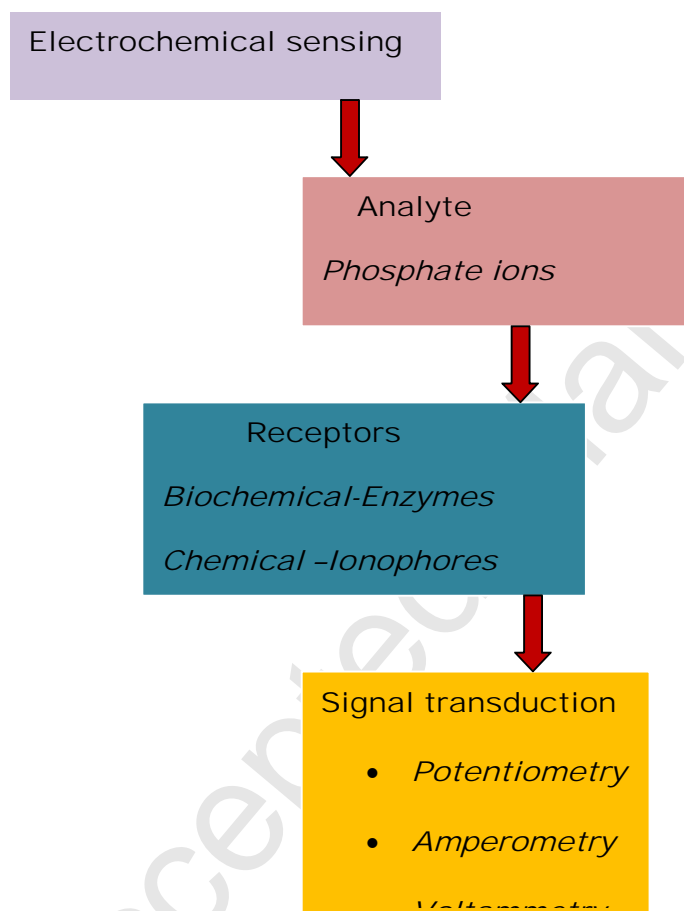
Figure 2 Recognition of phosphate based on supramolecular interactions

Figure 3 Schematic diagram of phosphate sensing based on the inhibition of the current due to the oxidation of glucose on Ni(OH)₂/NiOOH electrode in presence of phosphate ions

Figure 4 Schematic diagram of phosphate sensing based on the blocking of the ferrocyanide electron transfer kinetics when phosphate ions interact with Zr(IV) ions linked to a self assembled monolayer of mercaptopropionic acid.

Electro analysis of inorganic phosphate anions: A Review

Sheela Berchmans*, Touma B. Issa and Pritam Singh



Highlights

- Advantages of electrochemical sensing of phosphate
- Classification of electrochemical methods of sensing
- Ionophores for potentiometric sensing of phosphate
- Supramolecular based sensing of phosphate: Voltammetry and amperometry
- Unconventional and indirect methods of sensing phosphate

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Table 1 Analytical parameters for some potentiometric sensors for phosphate

| Sl.No | ISE (Active material) | Sensing parameters | Reference |
|-------|--|---|-----------|
| 1 | Co electrode | Dynamic linear range 10^{-4} - 10^{-2} mol L ⁻¹ , with slopes ranging from -35 to -50 mV/decade. | 40 |
| 2 | Co wire | Flow injection potentiometric(FIP) determinations of dihydrogenphosphate ($H_2PO_4^{2-}$) in fertilisers and waste waters at pH 5 have been carried out.Nernstian slope of -58.7 mV per decade was obtained in the concentration range 10^{-4} – 10^{-2} mol L ⁻¹ | 41 |
| 3 | Co micro electrode(10 μ m) | Dynamic linear range 10^{-5} to 10^{-1} mol L ⁻¹ Detection limit 7.5×10^{-5} mol L ⁻¹ . | 42 |
| 4 | oxidized cobalt metal electrodes | Linear dynamic range 10^{-5} - 10^{-2} mol L ⁻¹ at pH =4.0 ; (log $KH_2PO_4^1$ pot < -3). | 43 |
| 5 | Co wire electrode | - 38.0 ± 0.5 mV per decade in the range 5×10^{-3} – 10^{-5} mol L ⁻¹ at pH = 5.0 Detection limit 10^{-6} mol L ⁻¹ | 44 |
| 6 | Bis (p-chlorobenzyl) tin dichloride | 2.0×10^{-4} to 8.6×10^{-5} mol L ⁻¹ (This range is reported in the presence of different interfering anions) | 51 |
| 7 | Bis(terthiophene)-appended uranyl-salophen complex, comprising <i>N,N'</i> -bis[4-(5,2':5'',2''-terthiophen-3'-yl)salicylidene]-1,2-ethanediamine–uranyl complexes (TUS), as a monomer for the electrochemical polymerizations (poly-TUS) on glassy carbon surfaces to form functionalized conducting polymer (CP) films | The CP/poly-TUS sensor showed a linear range between 1.0×10^{-1} and $1.0 \times 10^{-4.5}$ mol L ⁻¹ with a near-Nernstian behavior (-30.4 mV/decade -1) at a pH of 8.2. The detection limit of the electrode was $10^{-5.0}$ mol L ⁻¹ and the response time was <10 s | 61 |
| 8 | 3-decyl-1,5,8-triazacyclodecane-2,4-dione) | Linear dynamic range 10^{-6} - 10^{-1} mol L ⁻¹ | 62 |
| 9 | A zwitterionic bis(guanidinium) ionophore bearing an anionic <i>clos</i> o-borane cluster | The lower detection limit for HPO_4^{2-} in an unbuffered solution is 8.7×10^{-8} mol L ⁻¹ | 64 |

Table 2 Analytical parameters for some enzymatic sensors for phosphate

| Sl.No | Enzyme electrode | Basis of measurement | Sensing parameters | Reference |
|-------|---|--|--|-----------|
| 1 | Immobilizing pyruvate oxidase (PyOD) on a screen-printed electrode. | The enzymatic generation of hydrogen peroxide (H ₂ O ₂) detected at +420mV vs Ag/AgCl | Response time <2 s short recovery time (2 min). The time required for one measurement using this phosphate biosensor was 4 min, which was faster than the time required using a commercial phosphate testing kit (10 min). The sensor has a linear range from 7.5 to 6.25 ×10 ⁻⁶ mol L ⁻¹ phosphate with a detection limit of 3.6 ×10 ⁻⁶ mol L ⁻¹ . Human salivary samples have been analysed for the phosphate content. | 4 |
| 2 | Poly(carbamoylsulphonate) (PCS)hydrogel immobilized pyruvate oxidase | Enzymatically generated H ₂ O ₂ monitored at +300mV versus phthalocyanin/carbon (PC) reference electrode | Rapid phosphate process control monitoring in an experimental sequencing batch reactor (SBR) system. The signal response time was 1 min with a detection limit of 5×10 ⁻³ mol L ⁻¹ | 20 |
| 3 | Pyruvate oxidase immobilized on a copolymer formed electrochemically with Os(bipy) ₂ pyCl modified pyrrole monomer and thiophene on a platinum black . | Detection of enzymatically generated H ₂ O ₂ (+0.40V versus Ag/AgCl) | Phosphate was measured similarly between 0.02 ×10 ⁻³ and 0.5×10 ⁻³ mol L ⁻¹ in the presence of pyruvate as co-substrate. The sensitivity of the sensor dropped to about 12% after 10 days. | 123 |
| 4 | Covalent immobilization of pyruvate oxidase (PyO) onto the nano-particle comprised poly- 5,2': 5,2''-terthiophene-3''-carboxylic acid, poly-TTCA (nano-CP) layers on a glassy carbon electrode | Detection of enzymatically generated H ₂ O ₂ (+0.40V versus Ag/AgCl) in a phosphate solution. | Dynamic linear range 1.0 ×10 ⁻⁶ mol L ⁻¹ to 100×10 ⁻⁶ mol L ⁻¹ and the detection limit was determined to be about 0.3 ×10 ⁻⁶ mol L ⁻¹ . The response time of the biosensors was about 6 s. | 124 |
| 5 | Immobilization of pyruvate oxidase (PyOx) on a polyion complex membrane | Detection of enzymatically generated H ₂ O ₂ | detection limit 0.2 ×10 ⁻⁶ mol L ⁻¹ of phosphoric acid | 125 |
| 6 | Maltose phosphorylase, acid phosphatase, glucose oxidase and mutarotase were coimmobilized on a regenerated cellulose membrane which was mounted on the tip of a platinum | A mperometric electrode for the detection of enzymatically formed hydrogen peroxide | Detection limit of 10 ⁻⁸ mol L ⁻¹ was obtained Dynamic range 0.1-1 ×10 ⁻⁶ mol L ⁻¹ , Relevant for the monitoring of water pollution. | 130 |
| 7 | Maltose phosphorylase (MP) from recombinant <i>Escherichia coli</i> immobilized on a planar interdigitated electrode by cross-linking with saturated glutaraldehyde (GA) vapour in the presence of bovine serum albumin | Conductometric biosensor | Temperature stability 20 °C to 50 ° Response time 10 s. The sensor has two linear ranges, one is from 1.0 to 20 ×10 ⁻⁶ mol | 134 |

| | | | | |
|----|---|--|--|-----|
| | (BSA). | | <p>L^{-1} phosphate with a detection limit of $1.0 \times 10^{-6} \text{ mol L}^{-1}$, and the other is between 20 and $400 \times 10^{-6} \text{ mol L}^{-1}$ phosphate.</p> <p>Storage life in citrate buffer was two months. No obvious interference from other anionic species like SO_4^{2-}, Cl^-, NO_3^-, NO_2^- and HCO_3^- was detected. The biosensor is suitable for use in real water samples</p> | |
| 8 | A potato (<i>Solanum tuberosum</i>) tissue slice immobilized glucose oxidase coupled with a Clark oxygen electrode. | Measurement is based on the inhibition by phosphate of potato acid phosphatase catalyzed glucose & phosphate | Lower detection limit $2.5 \times 10^{-5} \text{ mol L}^{-1}$; Sensor is stable for 28 days or 300 assays | 135 |
| 9 | The co-immobilization of polyphenol oxidase and Alkaline phosphatase leads to a bienzyme electrode for the determination of phosphate based on the inhibition effect of hydrolysis by phosphate. | Sensing is based on the detection of the enzymically generated oquinone at -0.2 V. | The sensitivity and detection limit of the biosensor for phosphate were $1.27 \text{ mAM}^{-1} \text{ cm}^{-2}$ and $2 \times 10^{-6} \text{ mol L}^{-1}$ | 140 |
| 10 | Phosphate-binding protein (PBP) from <i>Escherichia coli</i> . PBP was immobilized on a sheet of nitrocellulose membrane by cross-linking and the membrane potential of the immobilized PBP was measured. | | The response was selective to phosphate among other anions. Under optimum conditions $0.1-1.5 \times 10^{-3} \text{ mol L}^{-1}$ phosphate can be determined with this system. | 142 |

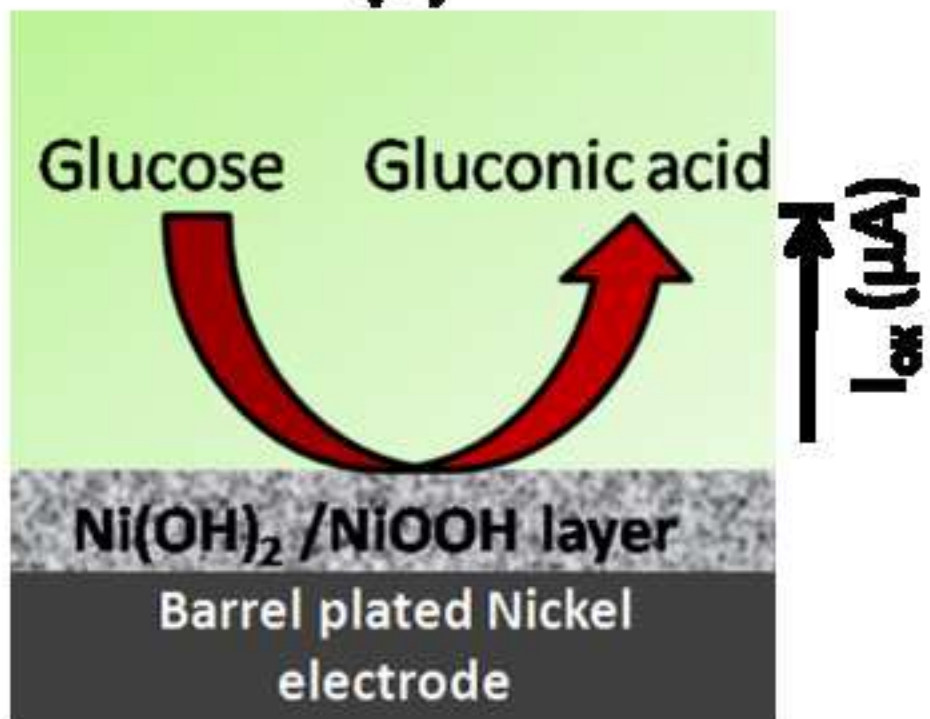


R - Redox reporters (Ferrocene, Cobaltocene...etc)
B - Binding site (Tetra alkyl ammonium group, Porphyrin, Pyrrole, Urea, Thio Urea ...etc)

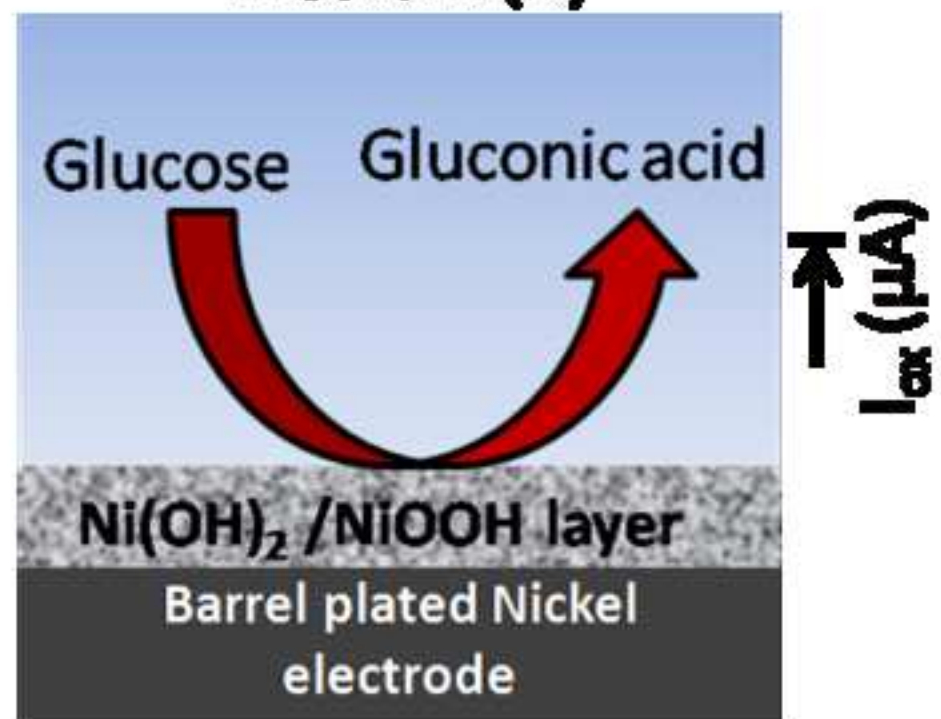
Electroanalysis

- **Cyclic voltammetry**
- **Differential pulse voltammetry**
- **Amperometry**

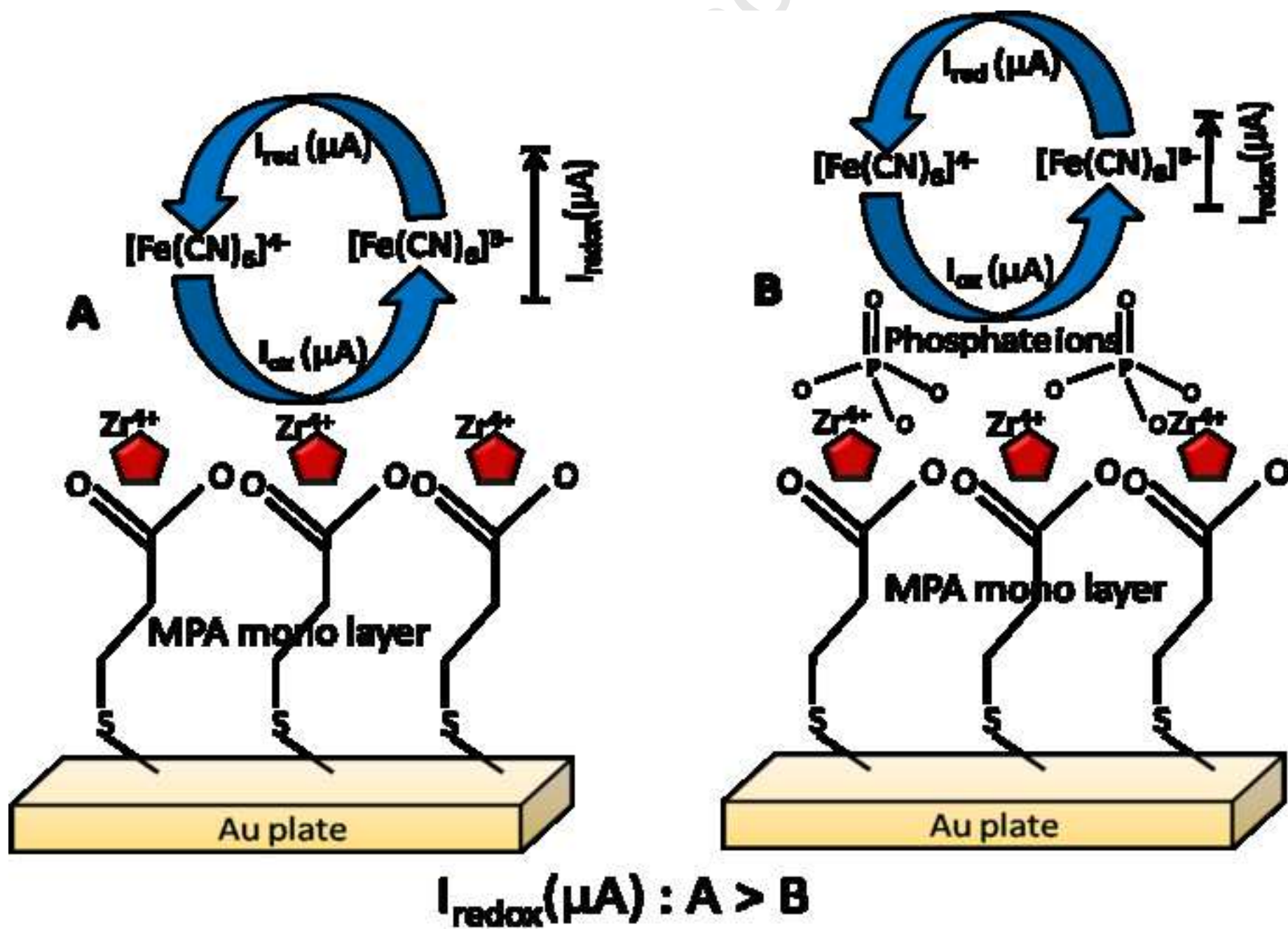
Phosphate free medium (A)



Phosphate containing medium (B)

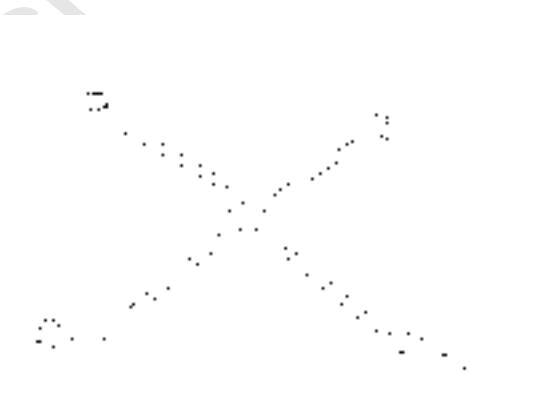


$I_{ox}(\mu A) : A > B$





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Digit 8 with bounding box



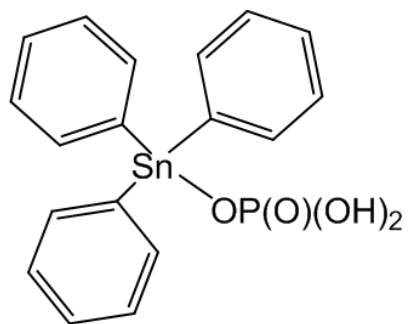
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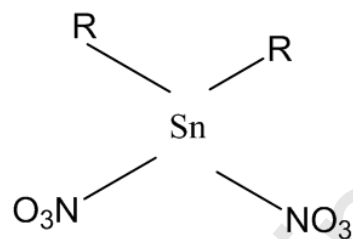
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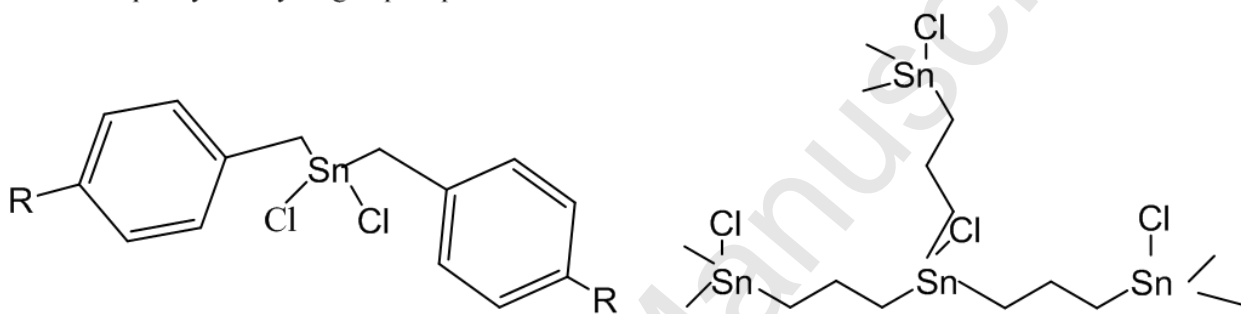
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Triphenyl tin hydrogen phosphate

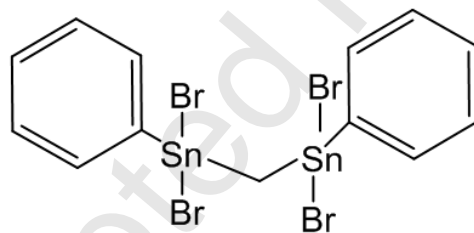


Dialkyl tin dinitrate ionophores



Dibenzyl chloride

Multidentate carrier with four tin centres



Distannyl derivative





