

**Plant Mechanisms Contributing
to Acid Impairment of Nodulation of
Medicago murex and *Medicago sativa*
by *Sinorhizobium medicae***

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I declare that this thesis is my own account of my research and contains as its main content work which has not previously been submitted for a degree at any tertiary education institution.

Yvonne Cheng

Cause is effect concealed, and effect is cause revealed.

The Aghori Vimalananda

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PUBLICATIONS ARISING FROM THE THESIS

Cheng Y, Watkin ELJ, O'Hara GW, Howieson JG. 2002. *Medicago sativa* and *Medicago murex* differ in the nodulation response to soil acidity. *Plant and Soil* **238**: 31-39.

Cheng Y, Howieson JG, O'Hara GW, Watkin ELJ, Souche G, Jaillard B, Hinsinger P. 2003. Proton release by roots of *Medicago murex* and *Medicago sativa* growing in acidic conditions, and implications for rhizosphere pH changes and nodulation at low pH. Accepted for publication in *Soil Biology and Biochemistry*.

SUMMARY

The widespread sowing of the perennial forage legume *Medicago sativa* to lower groundwater tables in south-western Australia is limited as many soils targeted for its use are too acidic ($\text{pH}_{\text{CaCl}_2} < 5$) for consistent nodulation with *Sinorhizobium* spp. The annual medic *M. murex* grows and nodulates well in these acidic soils, but it cannot fill the niche of *M. sativa* in lowering groundwater tables. The differential ability of *M. murex* and *M. sativa* to nodulate in acid soils provided the opportunity to compare the nodulation responses between the two species and to identify the mechanisms contributing to the poor nodulation of *M. sativa* in soil of low pH.

An initial glasshouse experiment compared the nodulation of *M. murex* cv. Zodiac and *M. sativa* cv. Aquarius with *S. medicae* strains WSM419 and CC169. Subsequent glasshouse and laboratory experiments used only the more acid-tolerant *S. medicae* strain WSM419. In the glasshouse in soil of $\text{pH}_{\text{CaCl}_2}$ 4.3, the uppermost nodule on both *M. murex* and *M. sativa* formed at 4-5 cm below the hypocotyl, but the nodules on *M. sativa* formed almost 4 weeks later than those on *M. murex*. The difference in nodulation response between *M. murex* and *M. sativa* was related to numbers of *S. medicae* in the rhizosphere. After 24 d growth in soil of $\text{pH}_{\text{CaCl}_2}$ 4.3, there were 100-fold higher numbers of *S. medicae* WSM419 associated with the roots of *M. murex* than *M. sativa*. This difference in rhizobial numbers was not due to differences in root growth as there were similar rates of root elongation in *M. murex* and *M. sativa*, or differences in the root products released as root exudates of *M. murex* and *M. sativa* produced at low pH had no significant effect on the growth of *S. medicae*.

Using a 'root mat' approach on soil disks of $\text{pH}_{\text{CaCl}_2}$ 4.49, *M. sativa* acidified its rhizosphere by approximately 0.2-0.4 pH-units within 4 d, while *M. murex* did not acidify its rhizosphere. Rates of H^+ release were higher from *M. sativa* than from *M. murex*. Using videodensitometry with agarose of pH 4.5, mature parts of the tap-root of both species exuded OH^- ions, but this was approximately 2-times higher in *M. murex* than in *M. sativa*. Consequently, young parts of the *M. sativa* rhizosphere were more acidic than that of *M. murex*. The higher rate of acidification by the roots of *M. sativa* made its rhizosphere less favourable for the survival and growth of *S. medicae*.

Root hair development was initially similar for both *M. murex* and *M. sativa*. However by 7 d after sowing in soil of $\text{pH}_{\text{CaCl}_2}$ 4.3, the density of root hairs on *M. murex* increased to 37 root hairs mm^{-1} root, while the density of root hairs on *M. sativa* decreased to 20 root hairs mm^{-1} root. Due to higher root hair density, the roots of *M. murex* provided a greater surface area for the attachment and colonisation of *S. medicae* compared to the roots of *M. sativa*. Indeed, confocal laser scanning microscopy at 7 d after sowing showed there were larger populations of a green fluorescent protein-marked transconjugant of *S. medicae* WSM419 colonised at 4-5 cm below the hypocotyl on the root of *M. murex* (3.28 pixel intensity units) compared to *M. sativa* (1.78 pixel intensity units). The smaller population of *S. medicae* colonised on the *M. sativa* root resulted in the observed delay in nodule development in *M. sativa* compared to *M. murex*.

Two plant mechanisms contributed to the greater numbers of *S. medicae* in the *M. murex* rhizosphere compared to *M. sativa* rhizosphere when plants were grown in an acidic soil: (1) roots of *M. murex* had a higher density of root hairs, and thus provided a larger root surface area for the growth and

colonisation of *S. medicae* than *M. sativa*, and (2) roots of *M. murex* acidified the rhizosphere less, and thus provided more favourable conditions for the growth and colonisation of *S. medicae* than the rhizosphere of *M. sativa*.

Models explaining the different nodulation responses between *M. murex* and *M. sativa* in soil of $\text{pH}_{\text{CaCl}_2}$ 4.3 and 7.0 are presented.

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A NOTE ABOUT *SINORHIZOBIUM MELILOTI* AND *S. MEDICAE* IN THE THESIS

The rhizobial strains used in the experiments described in this thesis are referred to as *Sinorhizobium medicae* based on their 16 rDNA sequence and host nodulation (G Garau, pers. comm., March 2003). However, *S. meliloti* and *Rhizobium meliloti* described in previously published literature are referred to as *S. meliloti*.