



## RESEARCH REPOSITORY

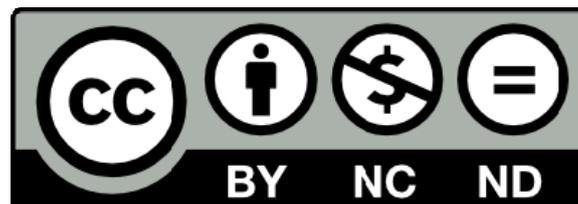
*This is the author's final version of the work, as accepted for publication following peer review but without the publisher's layout or pagination.*

*The definitive version is available at:*

<https://doi.org/10.1016/j.ecoleng.2017.05.027>

**Walden, L.L., Harper, R.J., Sochacki, S.J., Montagu, K.D., Wocheslander, R., Clarke, M., Ritson, P., Emms, J., Davoren, C.W., Mowat, D., Smith, A.P. and Gupta, V.V.S.R. (2017) Mitigation of carbon using Atriplex nummularia revegetation. Ecological Engineering, 106 . pp. 253-262.**

<http://researchrepository.murdoch.edu.au/37324/>



Crown Copyright © 2017.

# Mitigation of carbon using *Atriplex nummularia* revegetation

L.L.Walden<sup>a</sup>, R.J.Harper<sup>a</sup>, Sochacki<sup>a</sup>, K.D. Montagu<sup>b,c</sup>, R.Wocheslander<sup>a</sup>, M.Clarke<sup>d</sup>, P.Ritson<sup>e</sup>,  
J.Emms<sup>f</sup>, C.W.Davoren<sup>g</sup>, D.Mowat<sup>g</sup>, A.P.Smith<sup>g</sup>, V.V.S.RGupta<sup>g</sup>

<sup>a</sup>School of Veterinary and Life Sciences, Murdoch University, 90 South Street, Murdoch, WA 6150  
Australia

<sup>b</sup>State Forests of New South Wales, PO Box 100, Beecroft, NSW 2119, Australia

<sup>c</sup>Colo Consulting, Winmalee NSW, 2777 Australia

<sup>d</sup>Department of Agriculture and Food Western Australia, 20 Gregory St, Geraldton, WA, 6530  
Australia

<sup>e</sup>FarmWoods Consulting, Perth, WA, 6849 Australia

<sup>f</sup>SARDI, Hartley Grove, Urrbrae SA, 5064 Australia

<sup>g</sup>CSIRO Land and Water, Private Mail Bag 2, Glen Osmond, SA, 5064, Australia

## Abstract

The use of abandoned or marginally productive land to mitigate greenhouse gas emissions may avoid competition with food and water production. *Atriplex nummularia* Lindl. is a perennial shrub commonly established for livestock forage on saline land, however, its potential for carbon mitigation has not been systematically evaluated. Similarly, although revegetation is an allowable activity to mitigate carbon within Article 3.4 of the United Nations Framework Convention on Climate Change's Kyoto Protocol, there is a paucity of information on rates of carbon mitigation in soils and biomass through this mechanism. For six sites where *A. nummularia* had been established across southern

Australia four were used to assess changes in soil carbon storage and four were used to develop biomass carbon sequestration estimates. A generalised allometric equation for above and below ground biomass was developed, with a simple crown volume index explaining 81% of the variation in total biomass. There were no significant differences in soil organic carbon storage to 0.3 m or 2 m depth compared to existing agricultural land-use. Between 2.2 and 8.3 Mg C ha<sup>-1</sup> or 0.2–0.6 Mg C ha<sup>-1</sup> yr<sup>-1</sup> was sequestered in above and below ground biomass and this translates to potential total sequestration of 1.1–3.6 Tg C yr<sup>-1</sup> on saline land across Australia. Carbon income and forage grazing may thus provide a means to finance the stabilization of compromised land.

**Keywords:** Carbon sequestration; Co-benefits; Soil carbon; Desertification; Salinity; Land restoration

## 1. Introduction

Carbon dioxide (CO<sub>2</sub>) levels have increased by 40% since pre-industrial times, and as a consequence global surface and ocean surface temperatures show a warming of 0.85 °C over the period of 1880–2012 (IPCC 2013). The increase in atmospheric CO<sub>2</sub> has mainly occurred through the combustion of fossil fuels and the conversion of forests to agriculture (Battle et al., 2000; Le Quéré et al., 2013). Across different emission scenarios the mean global surface temperature is projected to increase by between 1.5 °C and 3 °C relative to 1850–1900 by the end of the 21<sup>st</sup> century (IPCC 2013).

A diverse range of policies, instruments and technologies are required to create a significant reduction in greenhouse gas (GHG) emissions and thus reduce the rate of temperature increase (IPCC 2014). The land sector both emits GHGs but also plays a central role in their mitigation (Bustamante et al., 2014; Smith et al., 2014). There are three main land sector approaches to mitigation: (1) the production of bioenergy from biomass to offset fossil fuel use (Chum et al., 2011), (2) carbon sequestration which is the transfer of atmospheric carbon dioxide into long-lived carbon pools in soils and plants (Lal 2008) and (3) the reduction in emissions from sources including deforestation, enteric fermentation in domestic animals and nitrogen fertilizers (Smith et al., 2014). Most research on land-based carbon mitigation has explored bio-sequestration via reforestation/afforestation, or storage in

soils (Smith et al., 2014). Revegetation, which is the establishment of vegetation that does not meet the definition of a forest, is an allowable activity under Article 3.4 of the Kyoto Protocol (UNFCCC 1997), but is characterized by a lack of literature that quantifies its potential contribution to climate change mitigation (Smith et al., 2014).

Carbon sequestration is often viewed as a win–win situation, providing environmental co-benefits while also mitigating atmospheric CO<sub>2</sub> (Harper et al., 2007; Bustamante et al., 2014). However, concerns have been raised with respect to reforestation and competition for land (Haberl et al., 2014), water (Jackson et al., 2005), reduced biodiversity (Lindenmayer et al., 2012) and the displacement of other land uses (Mitchell et al., 2012). This increases pressure on food security, and may be most pronounced in developing countries where food shortages are more prevalent (Foley et al., 2005; Smith et al., 2014).

Utilising abandoned or degraded land for carbon mitigation provides an opportunity to utilise land that has marginal potential for other uses (Silver et al., 2000; Sochacki et al., 2012; Wicke et al., 2011). Campbell et al. (2008) estimated that 385–472 million hectares of land is abandoned globally and suggested that potential mitigation on abandoned land is greatest in Australia, Brazil, and the United States. Prolonged inappropriate agricultural practices are a common cause of land degradation (Powlson et al., 2011) and Australia has large areas of land that are abandoned because of improper farming practices, such as over clearing and overgrazing (Pannell and Ewing 2006). For example, large expanses of land have become saline, reducing the productivity of the affected land (Lefroy and Stirzaker 1999). It is estimated that 5.7 Mha of land across Australia has salinized or is at risk of becoming saline, with this increasing to 17 Mha by 2050 (National Land and Water Resources Audit, 2001).

A form of rehabilitation of saline land is the establishment of halophytic saltbush species (e.g. *Atriplex* spp.) (Smith 2008). These species are typically planted in areas of marginal value for cropping or grazing, because of their resistance to drought and tolerance of poor soil conditions (de Araújo et al., 2006; Slavich et al., 1999). *Atriplex nummularia* Lindl. is commonly used as forage for grazing ruminants during times of feed shortages, as peak foliage production occurs in summer when

other feeds may be limited (Ben Salem et al., 2010). It follows that the majority of research involving this species has focussed on its use as livestock forage (Ben Salem et al., 2010; Glenn et al., 1998; Norman et al., 2004; Slavich et al., 1999).

The potential of *A. nummularia* established on abandoned and marginal agricultural land for carbon sequestration has not been formally published, despite the large amount of land potentially available for mitigation activities. The use of salt-land has however been considered for bioenergy production (Glenn et al., 1993; Sochacki et al., 2012; Wicke et al., 2011). Such a form of mitigation may be complementary to other forms of dryland carbon mitigation. Although mallee eucalypts are widely planted in Australian dryland carbon mitigation projects (Paul et al., 2015; Harper et al., 2017) these are not adapted to very saline soils.

Several assessments of the carbon mitigation potential of saltbush have remained in the grey literature (Issango et al., 2006; Montagu et al., 2003) or not been published at all. In this study, data from several of these studies and other unpublished data will be utilized, along with new field measurements. This study thus aims to explore the prospect of *A. nummularia* being used to sequester carbon on saline and poorly productive farmland in Australia by a) developing a more efficient method of estimating carbon in *A. nummularia* stands and in particular determining what explanatory variables contribute the most information to allometric relationships, b) producing an estimate of *A. nummularia* carbon sequestration potential at various sites across southern Australia, c) examining how carbon sequestered by *A. nummularia* is partitioned between soil organic carbon, and above and below ground biomass and d) examining factors such as planting density that affect the rates of sequestration by this species. This will allow policy makers, landholders and investors to determine whether carbon sequestration using this species is a viable carbon mitigation strategy, particularly in relation to other carbon mitigation approaches such as reforestation.

## **2. Methods**

### **2.1. Sampling sites**

Data were collected from six sites where *A. nummularia* had been established on farmland across southern Australia (Table 1, Fig. 1). These sites were considered representative of land that is marginal or degraded for typical agriculture in these areas. Across the six sites planting density varied from 500 to 2000 plants ha<sup>-1</sup> and stand age ranged from 4 to 13 years at the time of sampling (Table 1).

The six sites all fell within the Australian temperate climate zone, using the Köppen classification system, with wet and mild winters, hot dry summers (BOM 2017) and mean annual rainfall ranging from 248 to 404 mm yr<sup>-1</sup> (Table 1). The sites were grazed by sheep during late summer with edible dry matter (<2 mm stem size) removed. Plots were left to regrow and were measured in the following period, prior to grazing.

The sites were classified as saline or not saline based on soil measurements (Table 1). For Pithara, Three Springs and Wickepin a Geonics EM38 conductivity meter was used and for Monarto and Waikerie, salinity was measured on soil samples, and converted to EM38 equivalent values using the approach of McKenzie et al. (1989). No soil salinity data were available for Deniliquin.

### **2.2. Biomass sampling**

The carbon mitigation potential of a planting is often calculated using allometric equations. These equations relate explanatory values such as height, crown width, or a crown volume index (CVI) to the above, below, or total biomass of a given individual plant. The use of these equations allows carbon mitigation to be calculated without the need for destructive sampling.

At each site saltbush plants were destructively sampled as described by Snowdon et al. (2002). Plants were randomly selected across the dynamic range of plant sizes to ensure data were representative. Potential predictor variables of shrub height, crown height, length and width were measured and

applied in the development of allometric relationships. The number of plants sampled at each site is given in Table 1.

### **2.3. Above ground biomass**

At Deniliquin, Monarto, Waikerie and Wickepin the above ground biomass (AGB) was separated into components of stems and leaves (Fig. 2). Given that *A. nummularia* is mostly used as a forage shrub the biomass of foliage was not used in carbon calculations. The total fresh weight of each component was recorded and subsamples (0.5–0.7 kg) of each component were collected, dried at 70 °C to constant dry weight and total dry weight determined. At Pithara and Three Springs the above ground biomass was not separated into different components but weighed whole and then sub-sampled to estimate oven dry weight using the procedures above.

### **2.4. Below ground biomass**

At Wickepin, Monarto and Waikerie below ground biomass samples were taken by excavating soil by machine to 0.5 m and placing this on a sieving table overlaid with 50 mm square mesh as described by Ritson and Sochacki (2003) (Table 1). Samples were taken at the mid-point between adjacent shrub rows and the mid-point between shrubs within rows. All roots were collected to a minimum diameter limit of approximately 2 mm. Individual tap roots were excavated to a depth of 1 m. The roots were washed to remove any adhering soil and then dried, as described above, to determine the dry weight. Below ground biomass (BGB) was not measured at Pithara and Three Springs.

At Deniliquin, roots were sampled using a two stage method as described by Zerihun and Montagu (2004). The first stage involved soil coring for fine roots (<5 mm diameter) using a coring apparatus (Sochacki et al., 2007) with an internal diameter of 115 mm. Soil cores were collected to a depth of 1 m at increments of 0–0.1 m, 0.1–0.2 m, 0.2–0.3 m, 0.3–0.5 m and 0.5–1.0 m. Core samples were wet sieved to extract root material and dried at 70 °C. The second stage involved the excavation of sample plots as described above, to recover all roots  $\geq 5$  mm in diameter for all the sampled shrubs.

## **2.5. Soil organic carbon**

To assess changes in soil organic carbon (SOC) stocks at Wickepin, Deniliquin, Waikerie and Monarto soil samples were collected within treatment plots using the sampling methods described below.

### *2.5.1. Wickepin*

At Wickepin treatments consisted of replicated plots of two plant densities (500 and 2000 plants ha<sup>-1</sup>) as described by Sochacki et al. (2012). It was assumed that any changes in SOC would most likely be evident under high density treatments (2000 plants ha<sup>-1</sup>), thus only this treatment and the control were sampled. A coring apparatus (Sochacki et al., 2007) with an internal diameter of 115 mm was used to collect soil samples. Four control and six high density treatment plots were sampled. Four core samples were taken within each plot, one to a depth of 2 m, and three to a depth of 0.3 m. All cores were taken at a midpoint between the planting lines or approximately 1 m from the base of the saltbush shrub. The treatment plots were 40 × 40 m in size and cores were taken approximately 5 m inside the plot boundary to avoid edge effects. Depth intervals of coring were 0.1, 0.1–0.2, 0.2–0.3, 0.3–0.6, 0.6–0.9, 0.9–1.2, 1.2–1.5, 1.5–1.8 and 1.8–2.0 m.

### *2.5.2. Deniliquin*

For each plot at Deniliquin soil carbon concentration was measured on six bulked soil samples taken to a depth of 1 m. Four of these samples originated from bulking a subsample from the fine root sample cores. An additional two subsamples were taken from the soil bulk density samples. Samples were taken from the saltbush plots and control treatments were sampled from the adjacent pasture.

### *2.5.3. Waikerie and Monarto*

Soil samples were taken in depth increments of 0.05, 0.05–0.1, 0.1–0.3, 0.3–0.6, 0.6–1.0, 1.0–1.5 and 1.5–2.0 m adjacent to saltbush plants using a 4.5 cm diameter soil corer. A total of 6 samples were collected for each plant at distances of 0.2, 1 and 2 m from the base of the plant and there were three

replicates. Control samples were taken from pasture treatments at Monarto and from cropping treatments at Waikerie these being adjacent to the saltbush plots.

## **2.6. Soil analysis**

Soil samples were air-dried in a glasshouse prior to C analysis. Bulk density (BD) was determined for each depth interval by drying a subsample to 105 °C and correcting for moisture content to determine BD based on the mass-volume relationship.

Soil samples were sieved through a 2 mm sieve and the >2 mm fraction collected and weighed. The total SOC of the <2 mm fraction was determined by the Leco combustion method, LECO CNS 2000 analyser, method 6B3 (Rayment and Higginson 1992). Soils were also tested for the presence of calcium carbonate (McKenzie et al., 2000). Total SOC mass was calculated from the bulk density and carbon concentration, to express the total carbon content of the soil after taking into account the presence of any coarse (>2 mm) fractions using an equivalent soil mass (ESM) within a specified soil depth as described by Wendt and Hauser (2013).

## **2.7. Carbon analysis of *A. nummularia***

Carbon content of *A. nummularia* was determined from 8 plants at Wickepin, 9 plants from Waikerie, and 14 plants at Deniliquin. Sample shrubs were selected randomly from within the sample plots, separated into leaves and stems then dried at 70 °C to constant weight. The analysis for carbon content was undertaken at a commercial laboratory, using the Leco combustion method (Rayment and Higginson 1992). The carbon content of roots at Waikerie was also determined using the same analytical procedure.

## **2.8. Carbon isotopic analysis**

For soil from Deniliquin the stable carbon isotopic  $\delta^{13}\text{C}$  was determined on the fine root samples from 0 to 10 cm depth using a Europa Scientific automated nitrogen carbon analysis mass spectrometer as detailed in Krull and Bray (2005). This was conducted to determine the contribution of saltbush plants

to SOC pools. The relative proportions of fine roots of C4 plants (*A. nummularia*) and C3 plants (temperate pasture grasses) were estimated from the following equation (Ludlow et al., 1976):

$$\%C4 = ((\delta_s - \delta_{C3}) / (\delta_{C4} - \delta_{C3})) \times 100 \quad (1)$$

Where  $\delta_s$ ,  $\delta_{C3}$ , and  $\delta_{C4}$  are the  $\delta^{13}\text{C}$  values of, the fine root sample, average of the C3 grass, and average of the C4 *A. nummularia*.

The  $\delta^{13}\text{C}$  of soil samples from the 0–10 cm increment was analysed as detailed above. The relative proportions of derived soil organic carbon from of C4 plants (*A. nummularia*) and C3 plants (temperate pasture grasses) were estimated using Eq. (1), with  $\delta_s$  being the  $\delta^{13}\text{C}$  of the soil sample from either the saltbush or pasture plots.

## 2.9. Allometric equations

Allometric equations were developed using the biomass data from Wickepin, Monarto, Waikerie and Deniliquin (Fig. 3). Four models (linear with y-intercept, linear with y-intercept as 0, logarithmic and exponential) were fitted according to apparent trends within the data. Models were tested using the extra sum of square F-test to select the model with the best fit to the data. A fit index, standard error of estimate and coefficient of variation were also used to determine the model with the best fit to the data (Table 2). Normality assumptions were tested on all models. To test whether an all-inclusive (combined) model was possible, a dummy variable was used to test whether the difference in site-specific slopes was significant. If the slopes were not significantly different the sites were pooled to derive a combined model. Likewise, if no significant difference was found between saline and non-saline sites these two categories were combined. Models were fitted and tested using the software package R-Studio (R Core Team, 2014).

## 2.10. Estimation of carbon sequestration

The allometric relationships (Table 2) were applied to predict the amount of carbon sequestered per plant in the stems and roots and takes into account that leaves are grazed on an annual basis and thus don't represent a permanent component of the carbon pools. Carbon sequestered was calculated by

applying the allometric equations to stand data collected from permanent measurement plots to estimate the carbon storage in biomass at the different sites. Variation in stand density and age was considered when calculating sequestration per unit area over time.

Parameters to be applied as biomass predictor variables for *A. nummularia* were measured at all sites, these included shrub height, width and length. At Wickepin permanent measurement plots (0.04 ha) within larger treatment plots were assessed. Similarly, at Three Springs and Deniliquin 0.02 ha measurement plots were established while the Pithara site had 0.03 ha measurement plots. Given that the plots were of a known area and planting density, the data collected from these sites was then used to calculate carbon sequestration for each site.

The allometric equations were applied to the measured saltbush plants to estimate their total biomass. This was converted to carbon using the determined carbon content of components (Table 3) and carbon yield ( $\text{Mg ha}^{-1}$ ) was then calculated based on the planting density.

$$C = Db \times Pc \quad (2)$$

where:

*C* = Carbon, *Db* = dry biomass, *Pc* = carbon percentage of component.

## 2.11. Statistical analysis

Root to shoot ratios were analysed using one-way analysis of variance, with Tukey's honest significant difference post-hoc to determine site differences. This test was also used to compare the difference in carbon content (%) of the biomass components between the three sites. Normality assumptions were reviewed using Levene's test of equal variance and an examination of QQ plots. If data did not fulfil the normality assumptions, Welch's (1990) correction was used, as this test does not assume equal variance.

To test the differences in carbon sequestered across the different sites the Welch analysis of variance test, and Games-Howell post hoc test were used if the data did not satisfy homogeneity of variance

assumptions (Levene's test  $<0.05$ ). This method was also used to test the effect of stand age and density in the long-term data from Wickepin.

## **Results**

### **3.1. Saltbush partitioning**

There was some variation in the proportion each plant component contributed towards total plant biomass at each site (Fig. 2a). Saltbush at Monarto had the highest percentage of roots (40.1%) compared to Wickepin (25.6%), Waikerie (24.3%), and Deniliquin (15.3%). The stems made up the largest proportion of the total biomass of shrubs sampled at all the sites (51.8 – 71.7%), while the leaves made up the least (8.1 – 14.1%) with Wickepin having the highest percentage (14.1%) of leaves (Fig. 2a).

### **3.2. Root to shoot ratios**

The average root to shoot ratio at Monarto ( $0.70 \pm 0.10$ ) was significantly higher than that of Wickepin ( $0.39 \pm 0.06$ ) and Waikerie ( $0.37 \pm 0.11$ ) (Fig. 2b,  $P < 0.05$ ). The Deniliquin plants had the smallest mean root to shoot ratio ( $0.18 \pm 0.02$ ), and this was significantly less than the average root to shoot ratios at Wickepin ( $P < 0.05$ ) and Monarto ( $P < 0.05$ ). The mean root to shoot ratio across all sites was  $0.37 \pm 0.04$  (Fig. 2b).

### **3.3. Biomass allometric equations**

Allometric equations were developed for the combined above and below ground biomass referred to here as total biomass (TB), above ground biomass (AGB) and below ground biomass (BGB) using an array of field measurements. No significant difference was found between the slopes of these regressions for saline and non-saline sites, and therefore these data were combined. Shrub height, width, length, and height to crown base were measured to develop a crown volume index (CVI) (Sochacki et al., 2012). Five different shapes (hemisphere, cylinder, cone, cube, pyramidal) were tested against field biomass data to determine the best shape to calculate crown volume.

CVI provided the strongest relationship with TB, with 81% of the variation in TB explained by variation in CVI (Fig. 3c, Table 2). Height provided the weakest relationship of the three estimators ( $R^2 = 0.15$ , Fig. 3a). Crown diameter was only slightly less predictive than CVI ( $R^2 = 0.68$ , Fig. 3b). As the CVI of a plant increases so do the relative variance in TB estimates.

### **3.4. Carbon content of *A. nummularia***

For the three sites the carbon content of the plant components was similar, although the carbon content of the stems at Wickepin (49.2%) was significantly higher than Waikerie (47.3%,  $P < 0.001$ ). The leaf component had the lowest carbon content (Table 3) at all three sites, however the values of 36.5 and 38.8% at Waikerie and Wickepin were not significantly different ( $P > 0.05$ ). The mean carbon content of the stems was significantly higher than the carbon content of the leaves ( $37.6 \pm 0.64\%$ ,  $P < 0.001$ ), and significantly higher carbon than the roots ( $45.7 \pm 0.17\%$ ,  $P < 0.001$ ).

### **3.5. Soil carbon storage**

There were no significant differences in soil carbon storage between saltbush and control treatments at Wickepin, Deniliquin Waikerie or Monarto (Table 4). There was also no significant difference found at any of the depth intervals sampled. Carbon isotope analysis at Deniliquin indicated that the relative proportions of soil carbon (0–0.1 m) of C4 origin were 4.5% and 0% respectively in saltbush and pasture plots.

### **3.6. Carbon sequestration**

Using Eq. (2) and converting the estimates of biomass yield on an area basis, the amount of carbon sequestered at the four saline sites ranged from 2.2–8.3 Mg C ha<sup>-1</sup>, or 0.2–0.6 Mg C ha<sup>-1</sup> yr<sup>-1</sup> (Table 5).

At Wickepin, the effects of planting age and density on the rates of carbon sequestration were measured from 4 to 13 years after establishment. The cumulative amount of carbon sequestered across all densities increased from  $3.0 \pm 0.53$  Mg C ha<sup>-1</sup> when the planting was two years old, to  $8.3 \pm 0.73$  Mg C ha<sup>-1</sup> at 13 years of age (Fig. 4), with an apparent acceleration after eight years. Up

to eight years the higher density planting sequestered more carbon, but there was no difference in the cumulative sequestration in the subsequent period (Fig. 4).

## **4. Discussion**

### **4.1. Allometric relationships**

An allometric equation was derived for the estimation of total (above and below ground) biomass across a range of differing sites southern Australia. There was no difference between saline and non-saline sites (Fig. 3c). Interestingly, crown diameter was only slightly less predictive ( $R^2 = 0.68$ ) than CVI ( $R^2 = 0.81$ ) (Figs. 3b, c). The use of this predictor to remotely sense rates of carbon sequestration via aerial based techniques has been assessed by Liu et al. (2017). The combined TB allometric can be used for the development of broad scale carbon estimates from field data and also provides a means of quickly estimating stand biomass.

### **4.2. Carbon composition of *A. nummularia***

The partitioning and carbon content of *A. nummularia* has not been previously reported in the formal literature. The leaf components comprise a small proportion of the total biomass of the plant (8–14%, Fig. 2a), and also have the smallest carbon contents (36.5% & 38.8%, Table 3). These carbon values were consistent across both saline (Deniliquin, Wickepin) and non-saline (Waikerie) environments and may thus represent a biological trait of *A. nummularia*. The partitioning of the plant components between leaves, stems and roots did not vary markedly between sites, except for the plants sampled at Deniliquin. The majority of the biomass at this site was found in the stems (51.8 – 71.7%) and roots (15.3 – 40.1%) across all the sites, with the root: shoot ratio at most sites greater than the IPCC default value of 0.2, ranging from 0.37 to 0.70 for Waikerie, Wickepin and Monarto. Only Deniliquin had a value similar to the default value. The combined root to shoot ratio (0.37) across all sites was higher than previously reported for *A. nummularia* by Jones and Hodgkinson (1970), who reported ratios of 0.18 and 0.20 for saltbush occurring in natural communities. The

variation in root:shoot ratio may reflect site-specific characteristics, which could not be discerned in this study. Further research is required to extend these findings to other sites to determine possible relationships between root:shoot and site specific factors.

### **4.3. Carbon sequestration estimates**

The annual rate of carbon sequestered by *A. nummularia* of 0.20–0.64 Mg C ha<sup>-1</sup> yr<sup>-1</sup> across the sites (Table 5) was modest when compared to the average rate for reforestation with *Eucalyptus occidentalis* on similar land of 2.2 Mg C ha<sup>-1</sup> yr<sup>-1</sup> (Sochacki et al., 2012). The carbon sequestration rates reported here are similar to those of Issango et al. (2006) who estimated the carbon sequestered by grazed (0.2 Mg C ha<sup>-1</sup> yr<sup>-1</sup>) and ungrazed (0.3 Mg C ha<sup>-1</sup> yr<sup>-1</sup>) *Atriplex undulata* on saline land in Wickepin.

At Wickepin, *A. nummularia* sequestered 8.3 Mg C ha<sup>-1</sup> over a 13-year period, with no differences in sequestration with planting density after 8 years. The data reported here are limited to stands of age of 13 years, and therefore the evaluation of longer-term plantings and data are needed to provide an indication of the potential long-term carbon store of *A. nummularia*. Although Le Hou  rou (1992) suggested that saltbush growth may decline after 10 years there was no evidence of this at the Wickepin site; indeed growth accelerated in later years.

With an estimated 5.7 million ha of saline land in Australia potentially available for revegetation (National Land and Water Resources Audit, 2001), a first order estimate of carbon sequestration for Australia is 1.14–3.65 Tg C yr<sup>-1</sup>. Globally, 77 million ha of saline land may be similarly available for revegetation (Munns et al., 1999), with a possible annual rate of carbon sequestration of 15.4–49.3 Tg C yr<sup>-1</sup>. The yields reported here are smaller than the 5–10 Mg CO<sub>2</sub>-eq ha<sup>-1</sup> yr<sup>-1</sup> (1.4–2.7 Mg C ha<sup>-1</sup> yr<sup>-1</sup>) over 10 years for woody forage shrubs estimated from the National Carbon Accounting Toolbox (AGO 2005) by Monjardino et al. (2010) with consequent over-estimates of farm productivity improvements by using forage shrubs.

#### 4.4. Soil organic carbon

Changes in SOC levels in any cropping system will depend upon the quantity and quality of above and below ground plant C inputs, their decomposition rate and the management practices followed. Saltbush plants in the South Australian field experiments were grazed annually over a 4–8-week period at the end of summer (Descheemaeker et al., 2014; Revell et al., 2013), whereas at Wickepin saltbush was grazed at the end of summer effectively removing all leaf material. The lack of significant differences in SOC between saltbush treatments and agriculture controls may reflect the management regimes, the absence of sufficient litter fall and cycling of other necromass and high sampling errors due to a relatively low number of replicate soil cores taken per site.

There was no significant difference in the change in soil carbon storage to 0.3 m or indeed 2 m depth at Wickepin, and this is similar to what has been found for reforestation projects established on agricultural land in southern Australian environments (Harper et al., 2012; Hoogmoed et al., 2012). Also, the carbon isotopic analysis showed that only 4.5% of the soil carbon at Deniliquin originated from *A. nummularia*, suggesting that the turnover of carbon in these systems is low. There was no noticeable accumulation of litter at the sites, as they were all within grazing systems. Thus, it is recommended that SOC be omitted from carbon accounting of *A. nummularia* revegetation.

#### 4.5. Carbon accounting

For carbon accounting purposes, the carbon that would be sold in a trading scheme is that which represents a permanent increment (Penman et al., 2003) compared to the previous land-use. Carbon accounts for revegetation with *A. nummularia* will include stems and roots, whereas leaves and soft stem components will not form a part of the net carbon storage. Carbon sequestration in leaves at all the sites was calculated at less than  $0.1 \text{ Mg C ha}^{-1} \text{ yr}^{-1}$  (Fig. 2a), representing a minor component of the total carbon stocks. Grazing animals feed on the edible dry matter (leaves and stems <2 mm in diameter) (Andrew et al., 1976) and therefore carbon accounting for revegetation with *A. nummularia* will thus differ to reforestation projects, as not all of the biomass will be brought to account.

Although this study only examined carbon sequestration, revegetation with *A. nummularia* can also be valuable forage for summer grazing and land stabilization (Monjardino et al., 2010). Additionally, if saltbush is established on low value land, the opportunity cost or land rental associated with this conversion will be less than carbon mitigation activities (e.g. reforestation) on productive farmland (Townsend et al., 2012). The land conservation benefits and grazing potential from revegetation thus would need to be factored into any economic and sustainability analysis. There are potentially large areas of land that could be treated due to existing salinity or with limited other land-use options (Harper et al., 2007; Harper et al., 2017) and it may be possible to bundle the benefits from environmental service markets, such as carbon mitigation, together with grazing and salinity amelioration (Bennett et al., 2014) values. Importantly, such a system may represent one of the few economically viable options for saline land.

#### **4.6. Future research**

Ruminants that graze *A. nummularia* will also emit methane that will need to be included in any carbon accounting of saltbush grazing systems. The present version of the Australian National Carbon Accounting Toolbox for estimating carbon sequestration in different systems does not include estimates for woody shrubs such as saltbush (DEE, 2017). In addition, *A. nummularia* establishment may affect soil greenhouse gas emissions. *A. nummularia* is predominantly planted on saline, waterlogged soils and these soils are presumed to emit methane (Le Mer and Roger 2001), however the rates of methane emissions across the southern Australian sites are unknown as is the impact of reducing waterlogging through revegetation. Neither grazing nor soil emissions were investigated in this study and these are areas on which future research could focus.

### **5. Conclusions**

Further work is required to refine the estimates of carbon sequestration, at both site specific and aggregate scales for grazing shrub species on abandoned or compromised land. Nonetheless, this study found that revegetation with *A. nummularia* presents an opportunity to sequester carbon on land

that has marginal value for other uses. Apart from providing grazing value this will also provide environmental co-benefits such as stabilizing degraded farmland. Revegetating saline land with *A. nummularia* does not displace farm production but provides forage for grazing ruminants particularly during the typical end of summer feed shortage. Although there were modest amounts of carbon stored in the above and below ground biomass there are large areas of saline land and a first order estimate of the potential for carbon sequestration using *A. nummularia* on saline land in Australia is between 1.12–3.65 Tg C yr<sup>-1</sup>. The respective estimate globally across 77 million ha of abandoned saline land is 12.5–47.3 Tg C yr<sup>-1</sup>.

## **Acknowledgements**

We acknowledge funding from the Australian Government (Australian Joint Venture Agroforestry Program (Project CAL-8A), the Natural Heritage Trust Farm Forestry Program Project ‘Putting Trees in Their Place’ (983197), CRC Greenhouse Accounting, “Filling the Research Gap: National Soil Carbon Program” (Project 01203.077), the CRC Future Farm Industries and EverCrop, a GRDC-funded project. The latter also funded part of Bill Davoren’s salary. The Northern Agricultural Catchment Council (Western Australia), provided data. Murdoch University provided funds to support Lewis Walden’s research. Belinda Cale constructed Fig. 1 and Kate Düttmer, Piers Harper, Nick Saunders and David Giles assisted in the field and laboratory. We thank all landholders for enthusiastic participation in this research and Prof Ed. Barrett-Lennard, Prof. Richard Bell and Dr Surender Mann for useful discussions.

## **References**

- AGO, 2005. The National Carbon Accounting Toolbox and Data Viewer. Australian Greenhouse Office., Canberra
- Andrew, M., Noble, I., Lange, R., 1976. A non-destructive method for estimating the weight of forage on shrubs. Rangeland J. 1, 225–231.

- BOM, 2017. Key climate groups, Bureau of Meteorology climate zones. Australian Government Bureau of Meteorology, [www.bom.gov.au/iwk/climate/zones/map1.shtml](http://www.bom.gov.au/iwk/climate/zones/map1.shtml) (Accessed 10 April 2017).
- Battle, M., Bender, M.L., Tans, P.P., White, J.W.C., Ellis, J.T., Conway, T., Francey, R.J., 2000. Global carbon sinks and their variability inferred from atmospheric O<sub>2</sub> and <sup>13</sup>C. *Science* 287, 2467–2470.
- Ben Salem, H., Norman, H.C., Nefzaoui, A., Mayberry, D.E., Pearce, K.L., Revell, D.K., 2010. Potential use of oldman saltbush (*Atriplex nummularia* Lindl.) in sheep and goat feeding. *Small Ruminant Res.* 91, 13–28.
- Bennett, D., George, R., Silberstein, R., Ryder, A., Lambert, T., 2014. Changes in run-off and groundwater conditions under saltbush grazing systems: results of a ten year paired catchment study. *Hydrol. Water Resour. Symp.*, 1064–1071, Engineers Australia.
- Bustamante, M., Robledo-Abad, C., Harper, R., Mbow, C., Ravindranath, N.H., Sperling, F., Haberl, H., Siqueira Pinto, A., Smith, P., 2014. Co-benefits, trade-offs, barriers and policies for greenhouse gas mitigation in the Agriculture, Forestry and Other Land Use (AFOLU) sector. *Global Change Biol.* 20, 3270–3290.
- Campbell, J.E., Lobell, D.B., Genova, R.C., Field, C.B., 2008. The global potential of bioenergy on abandoned agriculture lands. *Environ. Sci. Technol.* 42, 5791–5794.
- Chum, H., Faaij, A., Moreira, J., Berndes, G., Dhamija, P., Dong, H., Gabrielle, B., Eng, A.G., Lucht, W., Mapako, M., Cerutti, O.M., McIntyre, T., Minowa, T., Pingoud, K., 2011. Bioenergy. In: Edenhofer, O., Pichs-Madruga, R., Sokona, Y., Seyboth, K., Matschoss, P., Kadner, S., Zwickel, T., Eickemeier, P., Hansen, G., Schlömer, S., von Stechow, C. (Eds.), *IPCC Special Report on Renewable Energy Sources and Climate Change Mitigation*. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.
- de Araújo, S.A., Silveira, J.A., Almeida, T.D., Rocha, I., Morais, D.L., Viégas, R.A., 2006. Salinity tolerance of halophyte *Atriplex nummularia* L. grown under increasing NaCl levels. *Revista Brasileira de Engenharia Agrícola e Ambiental* 10, 848–854.
- DEE, 2017. Land Sector Reporting. Overview of the Land Sector in Australia's National Inventory. Australian Government, Department of Environment and Energy, [www.environment.gov.au/climate-change/greenhouse-gasmeasurement/land-sector](http://www.environment.gov.au/climate-change/greenhouse-gasmeasurement/land-sector) (Accessed 10 April 2017).
- Descheemaeker, K., Smith, A., Robertson, M., Whitbread, A., Huth, N., Davoren, W., Emms, J., Llewellyn, R., 2014. Simulation of water-limited growth of the forage shrub saltbush (*Atriplex nummularia* Lindl.) in a low-rainfall environment of southern Australia. *Crop Pasture Sci.* 65, 1068–1083.
- Foley, J.A., DeFries, R., Asner, G.P., Barford, C., Bonan, G., Carpenter, S.R., Chapin, F.S., Coe, M.T., Daily, G.C., Gibbs, H.K., Helkowski, J.H., Holloway, T., Howard, E.A., Kucharik, C.J., Monfreda, C., Patz, J.A., Prentice, I.C., Ramankutty, N., Snyder, P.K., 2005. Global consequences of land use. *Science* 309, 570–574.
- Glenn, E., Squires, V., Olsen, M., Frye, R., 1993. Potential for carbon sequestration in the drylands. *Water Air Soil Pollut.* 70, 341–355.
- Glenn, E., Tanner, R., Miyamoto, S., Fitzsimmons, K., Boyer, J., 1998. Water use, productivity and forage quality of the halophyte *Atriplex nummularia* grown on saline waste water in a desert environment. *J. Arid Environ.* 38, 45–62.
- Haberl, H., Mbow, C., Deng, X., Irwin, E.G., Kerr, S., Kuemmerle, T., Mertz, O., Meyfroidt, P., Turner, I., 2014. *Finite Land Resources and Competition Rethinking Global Land Use in an Urban Era*. MIT Press, Cambridge.

- Harper, R.J., Beck, A.C., Ritson, P., Hill, M.J., Mitchell, C.D., Barrett, D.J., Smettem, K.R.J., Mann, S.S., 2007. The potential of greenhouse sinks to underwrite improved land management. *Ecol. Eng.* 29, 329–341.
- Harper, R.J., Okom, A.E.A., Stilwell, A.T., Tibbett, M., Dean, C., George, S.J., Sochacki, S.J., Mitchell, C.D., Mann, S.S., Dods, K., 2012. Reforesting degraded agricultural landscapes with Eucalypts: effects on soil carbon storage and soil fertility after 26 years. *Agric. Ecosyst. Environ.* 163, 3–13.
- Harper, R.J., Sochacki, S.J., McGrath, J.F., 2017. The development of reforestation options for dryland farmland in southwestern Australia: a review. *Southern For.* <http://dx.doi.org/10.2989/20702620.2016.1255417>.
- Hoogmoed, M., Cunningham, S.C., Thomson, J.R., Baker, P.J., Beringer, J., Cavagnaro, T.R., 2012. Does afforestation of pastures increase sequestration of soil carbon in Mediterranean climates? *Agric. Ecosyst. Environ.* 159, 176–183.
- IPCC, 2013. Climate Change. In: Stocker, T.F., Qin, D., Plattner, G.K., Tignor, M., Allen, S.K., Boschung, J., Nauels, A., Xia, Y., Bex, V., Midgley, P.M. (Eds.), *The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge, United Kingdom and New York.
- IPCC, 2014. Technical Summary. In: *Climate Change 2014: Mitigation of Climate Change. Working Group III Contribution to the IPCC 5th Assessment Report - Changes to the underlying Scientific/Technical Assessment*, Cambridge, United Kingdom and New York: Cambridge University Press.
- Issango, J.O., Bell, R.W., Waddell, B., Mann, S.S., Barrett-Lennard, E.G., 2006. Quantifying carbon sequestration on saltland pastures in south west Australia. In: Turner, N., Acuna, T. (Eds.), *Proceedings of the 13th Australian Agronomy Conference*. 10-14 September 2006 Perth. Western Australia.
- Jackson, R.B., Jobbágy, E.G., Avissar, R., Roy, S.B., Barrett, D.J., Cook, C.W., Farley, K.A., le Maitre, D.C., McCarl, B.A., Murray, B.C., 2005. Trading water for carbon with biological carbon sequestration. *Science* 310, 1944–1947.
- Jones, R., Hodgkinson, K., 1970. Root growth of rangeland chenopods: morphology and production of *Atriplex nummularia* and *Atriplex vesicaria*. Commonwealth Scientific and Industrial Research Organization Division of Plant Industry *The Biology of Atriplex*, 77–85.
- Krull, E.G., Bray, S.S., 2005. Assessment of vegetation change and landscape variability by using stable carbon isotopes of soil organic matter. *Aust. J. Bot.* 53, 651–661.
- Lal, R., 2008. Carbon sequestration. *Philosophical transactions of the royal society B. Biol. Sci.* 363, 815–830.
- Le Houérou, H.N., 1992. The role of saltbushes (*Atriplex* spp.) in arid land rehabilitation in the Mediterranean Basin: a review. *Agrofor. Syst.* 18, 107–148.
- Le Mer, J., Roger, P., 2001. Production, oxidation, emission and consumption of methane by soils: a review. *Eur. J. Soil Biol.* 37, 25–50.
- Le Quéré, C., Andres, R.J., Boden, T., Conway, T., Houghton, R.A., House, J.I., Marland, G., Peters, G.P., van der Werf, G.R., Ahlström, A., Andrew, R.M., Bopp, L., Canadell, J.G., Ciais, P., Doney, S.C., Enright, C., Friedlingstein, P., Huntingford, C., Jain, A.K., Jourdain, C., Kato, E., Keeling, R.F., Klein Goldewijk, K., Levis, S., Levy, P., Lomas, M., Poulter, B., Raupach, M.R., Schwinger, J., Sitch, S., Stocker, B.D., Viovy, N., Zaehle, S., Zeng, N., 2013. The global carbon budget 1959–2011. *Earth Syst. Sci. Data* 5, 165–185.
- Lefroy, E., Stirzaker, R., 1999. Agroforestry for water management in the cropping zone of southern Australia. *Agrofor. Syst.* 45, 277–302.

- Lindenmayer, D.B., Hulvey, K.B., Hobbs, R.J., Colyvan, M., Felton, A., Possingham, H., Steffen, W., Wilson, K., Youngentob, K., Gibbons, P., 2012. Avoiding bio-perversity from carbon sequestration solutions. *Conserv. Lett.* 5, 28–36.
- Liu, N., Harper, R.J., Handcock, R., Evans, B.J., Sochacki, S.J., Liu, S., Dell, B., Walden, L.L., 2017. Optimal timing for estimating carbon mitigation in revegetation of abandoned agricultural land with high spatial resolution remote sensing. *Remote Sensing* 9, 545, <http://dx.doi.org/10.3390/rs9060545>.
- Ludlow, M.M., Troughton, J.H., Jones, R.J., 1976. A technique for determining the proportion of C3 and C4 species in plant samples using stable natural isotope's of carbon. *J. Agric. Sci.* 87, 625–632.
- McKenzie, R.C., Chomistek, W., Clark, N.F., 1989. Conversion of electromagnetic inductance readings to saturated paste extract values in soils for different temperature, texture, and moisture conditions. *Can. J. Soil Sci.* 69, 25–32.
- McKenzie, N., Ryan, P., Fogarty, P., Wood, J., 2000. Sampling, measurement and analytical protocols for carbon estimation in soil, litter and coarse woody debris. Australian Greenhouse Office Canberra.
- Mitchell, C., Harper, R., Keenan, R., 2012. Status and prospects of carbon forestry in Australia. *Austr. For.* 75, 201–213.
- Monjardino, M., Revell, D., Pannell, D.J., 2010. The potential contribution of forage shrubs to economic returns and environmental management in Australian dryland agricultural systems. *Agric. Syst.* 103, 187–197.
- Montagu, K.D., Cowie, A.M., Rawson, A., Wilson, B.R., George, B.H., 2003. Carbon Sequestration Predictor for Land Use Change in Inland Areas of New South Wales. Background, User Notes, Assumptions and Preliminary Model Testing. Version 2.0. State Forests of New South Wales.
- Munns, R., Cramer, G.R., Ball, M.C., 1999. Interactions between rising CO<sub>2</sub>, soil salinity and plant growth. In: Mooney, H.A. (Ed.), *Carbon Dioxide and Environmental Stress*. Academic Press, London, pp. 139–167.
- National Land and Water Resources Audit, 2001. Dryland salinity in Australia: a summary of the National Land and Water Resources Audit's Australian Dryland Salinity Assessment 2000. National Land and Water Resources Audit, Canberra.
- Norman, H.C., Freind, C., Masters, D.G., Rintoul, A.J., Dynes, R.A., Williams, I.H., 2004. Variation within and between two saltbush species in plant composition and subsequent selection by sheep. *Austr. J. Agric. Res.* 55, 999–1007.
- Pannell, D.J., Ewing, M.A., 2006. Managing secondary dryland salinity: options and challenges. *Agric. Water Manage.* 80, 41–56.
- Paul, K.I., Roxburgh, S.H., de Ligt, R., Ritson, P., Brooksbank, K., Peck, A., Wildy, D.T., Mendham, D., Bennett, R., Bartle, J., Larmour, J.S., Raison, R.J., England, J.R., Clifford, D., 2015. Estimating temporal changes in carbon sequestration in plantings of mallee eucalypts: modelling improvements. *For. Ecol. Manage.* 335, 166–175.
- Penman, J., Gytarsky, M., Hiraishi, T., Krug, T., Kruger, D., Pipatti, R., Buendia, L., Miwa, K., Ngara, T., Tanabe, K., Wagner, F. (Eds.), 2003. *Hayama, Japan : Intergovernmental Panel on Climate Change. National Greenhouse Gas Inventories Programme.*
- Powlson, D.S., Whitmore, A.P., Goulding, K.W.T., 2011. Soil carbon sequestration to mitigate climate change: a critical re-examination to identify the true and the false. *Eur. J. Soil Sci.* 62, 42–55.
- R Core Team, 2014. *R: A language and environment for statistical computing.* R Foundation for Statistical Computing., Vienna.

- Rayment, G.E., Higginson, F.R., 1992. Australian Laboratory Handbook of Soil and Water Chemical Methods. Inkata Press, Melbourne.
- Revell, D., Norman, H., Vercoe, P., Phillips, N., Toovey, A., Bickell, S., Hulm, E., Hughes, S., Emms, J., 2013. Australian perennial shrub species add value to the feed base of grazing livestock in low-to medium-rainfall zones. *Anim. Prod. Sci.* 53, 1221–1230.
- Ritson, P., Sochacki, S., 2003. Measurement and prediction of biomass and carbon content of *Pinus pinaster* trees in farm forestry plantations, south-western Australia. *For. Ecol. Manage.* 175, 103–117.
- Silver, W.L., Ostertag, R., Lugo, A.E., 2000. The potential for carbon sequestration through reforestation of abandoned tropical agricultural and pasture lands. *Restor. Ecol.* 8, 394–407.
- Slavich, P.G., Smith, K.S., Tyerman, S.D., Walker, G.R., 1999. Water use of grazed salt bush plantations with saline watertable. *Agric. Water Manage.* 39, 169–185
- Smith, P., Bustamante, M., Ahammad, H., Clark, H., Dong, H., Elsiddig, E.A., Haberl, H., Harper, R., House, J., Jafari, M., Maser, O., Mbow, C., Ravindranath, N.H., Rice, C.E., Robledo Abad, C., Romanovskaya, A., Sperling, F., Tubiello, F.N., 2014. Agriculture, Forestry and Other Land Use (AFOLU). In: Edenhofer, O., Pichs-Madruga, R., Sokona, Y., Farahani, E., Kadner, S., Seyboth, K., Adler, A., Baum, I., Brunner, S., Eickemeier, P., Kriemann, B., Savolainen, J., Schlömer, S., von Stechow, C., Zwickel, T., Minx, J.C. (Eds.), *Climate Change 2014: Mitigation of Climate Change Contribution of Working Group III to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA, pp. 811–922.
- Smith, F.P., 2008. Who's planting what, where and why—and who's paying?: An analysis of farmland revegetation in the central wheatbelt of Western Australia. *Landscape Urban Plann.* 86, 66–78.
- Snowdon, P., Keith, H., Raison, R.J., 2002. Protocol for sampling tree and stand biomass: Australian Greenhouse Office Parkes. ACT.
- Sochacki, S., Ritson, P., Brand, B., 2007. A specialised soil corer for sampling tree roots. *Aust. J. Soil Res.* 45, 111–117.
- Sochacki, S.J., Harper, R.J., Smettem, K.R., 2012. Bio-mitigation of carbon following afforestation of abandoned salinized farmland. *GCB Bioenergy* 4, 193–201.
- Townsend, P.V., Harper, R.J., Brennan, P.D., Dean, C., Wu, S., Smettem, K.R.J., Cook, S.E., 2012. Multiple environmental services as an opportunity for watershed restoration. *For. Policy Econ.* 17, 45–58.
- UNFCCC, 1997. Kyoto Protocol to the United Nations Framework Convention on Climate Change. United Nations Framework Convention on Climate Change.
- Welch, W.J., 1990. Construction of permutation tests. *J. Am. Stat. Assoc.* 85, 693–698.
- Wendt, J., Hauser, S., 2013. An equivalent soil mass procedure for monitoring soil organic carbon in multiple soil layers. *Eur. J. Soil Sci.* 64, 58–65.
- Wicke, B., Smeets, E., Dornburg, V., Vashev, B., Gaiser, T., Turkenburg, W., Faaij, A., 2011. The global technical and economic potential of bioenergy from salt-affected soils. *Energy Environ. Sci.* 4, 2669–2681.
- Zerihun, A., Montagu, K.D., 2004. Belowground to aboveground biomass ratio and vertical root distribution responses of mature *Pinus radiata* stands to phosphorus fertilization at planting. *Can. J. For. Res.* 34, 1883–1894.

Table 1. Summary of environmental features and sampling regime at each of the sites.

| Site          | Location     |               | Planting Density<br>(plants ha <sup>-1</sup> ) | Rainfall<br>(mm) | Salinity<br>(mS m <sup>-1</sup> ) | Age of plantation<br>(years) | Number of plants sampled |     | Sampling methods |                       |
|---------------|--------------|---------------|--|------------------|-----------------------------------|------------------------------|--------------------------|-----|------------------|-----------------------|
|               | Lat.<br>(°S) | Long.<br>(°E) |  |                  |                                   |                              | AGB                      | BGB | AGB              | BGB                   |
| Deniliquin    | 35.293       | 144.452       | 1250   | 337              | –                                 | 5                            | 14                       | 14  | F.H              | Excavation<br>+ Cores |
| Monarto       | 35.717       | 139.848       | 1111   | 368              | 33                                | 4                            | 12                       | 6   | F.H              | Excavation            |
| Pithara       | 30.363       | 116.729       | 1411   | 315              | 192                               | 11                           | 14                       | 0   | F.H              | –                     |
| Three Springs | 29.622       | 115.649       | 892  | 382              | 288                               | 13                           | 12                       | 0   | F.H              | –                     |
| Waikerie      | 32.433       | 117.394       | 635  | 248              | 23                                | 4                            | 9                        | 9   | F.H              | Excavation            |
| Wickepin      | 32.433       | 117.394       | 500, 2000                                      | 404              | 190                               | 13                           | 54                       | 22  | F.H              | Excavation            |

Salinity measured with EM38; AGB: Above Ground Biomass; BGB: Below Ground Biomass; F.H: Full harvest.

Table 2. Allometric equations for the prediction of above ground biomass (AGB), below ground biomass (BGB) and total biomass (TB) in *Atriplex nummularia*, including a range of goodness-of-fit statistics. These were based on the crown volume index (CVI), diameter (d) and height (h) of the plant.

| n          | Allometric equation | R <sup>2</sup>          | FI   | SE   | CV (%) |
|------------|---------------------|-------------------------|------|------|--------|
| <i>AGB</i> |                     |                         |      |      |        |
| CVI        | 51                  | AGB = 3.11 * CVI + 2.53 | 0.79 | 0.74 | 33.9   |
| <i>BGB</i> |                     |                         |      |      |        |
| CVI        | 51                  | BGB = 0.80 * CVI + 1.12 | 0.59 | 0.59 | 37.3   |
| <i>TB</i>  |                     |                         |      |      |        |
| Height     | 51                  | TB = 12.0 * h - 7.55    | 0.15 | 0.15 | 56.6   |
| Diameter   | 51                  | TB = 8.54 * d           | 0.68 | 0.68 | 34.8   |
| CVI        | 51                  | TB = 4.67 * CVI + 3.7   | 0.81 | 0.81 | 26.4   |

n is the number of samples, R<sup>2</sup> the coefficient of determination, FI is fit index, SE standard error of estimate and CV (%) is the coefficient of variation.

Table 3. Carbon content (%) of the plant components with standard errors.

| Site             | Carbon content (%) |                    |                    |
|------------------|--------------------|--------------------|--------------------|
|                  | AGB                | BGB                |                    |
|                  | Leaf               | Stem               | Root               |
| Deniliquin       | 37.3 ( $\pm 0.3$ ) | 46.7 ( $\pm 0.3$ ) | 46.3 ( $\pm 0.2$ ) |
| Waikerie         | 36.5 ( $\pm 0.3$ ) | 47.3 ( $\pm 0.1$ ) | 45.7 ( $\pm 0.2$ ) |
| Wickepin         | 38.8 ( $\pm 1.1$ ) | 49.2 ( $\pm 0.2$ ) | N.A                |
| Combined average | 37.3 ( $\pm 0.6$ ) | 47.7 ( $\pm 0.3$ ) | 46.0 ( $\pm 0.2$ ) |

AGB: Above Ground Biomass; BGB: Below Ground Biomass.

Table 4. Soil carbon storage (t C ha<sup>-1</sup>) for different depth intervals (0-0.3, 0-3-2.0, 0-1.0 m). None of the paired comparisons between saltbush and pasture were significantly different.

| Site       | 0-0.3 m |          | 0.3-2.0 m |          | 0-1.0 m |          |
|------------|---------|----------|-----------|----------|---------|----------|
|            | Pasture | Saltbush | Pasture   | Saltbush | Pasture | Saltbush |
| Deniliquin | -       | -        | -         | -        | 64.9    | 71.2     |
| Monarto    | 43.44   | 45.39    | 31.08     | 35.16    | -       | -        |
| Waikerie   | 15.39   | 14.65    | 35.69     | 38.14    | -       | -        |
| Wickepin   | 27.91   | 27.67    | 22        | 22.52    | -       | -        |

- : indicates where data were not available for the given sampling depth.

Table 5. Estimates of carbon sequestration for the four saline sites using the AGB and BGB allometric equations. Standard errors are reported in brackets. Plot-scale estimates were not available for the non-saline sites at Waikerie or Monarto.

| Site          | Age | Carbon sequestration |                                       |
|---------------|-----|----------------------|---------------------------------------|
|               | yr  | t C ha <sup>-1</sup> | t C ha <sup>-1</sup> yr <sup>-1</sup> |
| Deniliquin    | 5   | 2.16(±0.10)          | 0.43(±0.02)                           |
| Pithara       | 11  | 2.27(±0.53)          | 0.20(±0.05)                           |
| Three Springs | 13  | 4.79(±1.07)          | 0.37(±0.08)                           |
| Wickepin      | 13  | 8.33(±0.73)          | 0.64(±0.06)                           |

Fig. 1. Map showing position of sites sampled across southern Australia. Inset maps show more detailed positioning of sites with mean annual rainfall (mm/yr) isohyets. The Wickepin, Monarto, Waikerie and Deniliquin sites were used to develop the allometric model.

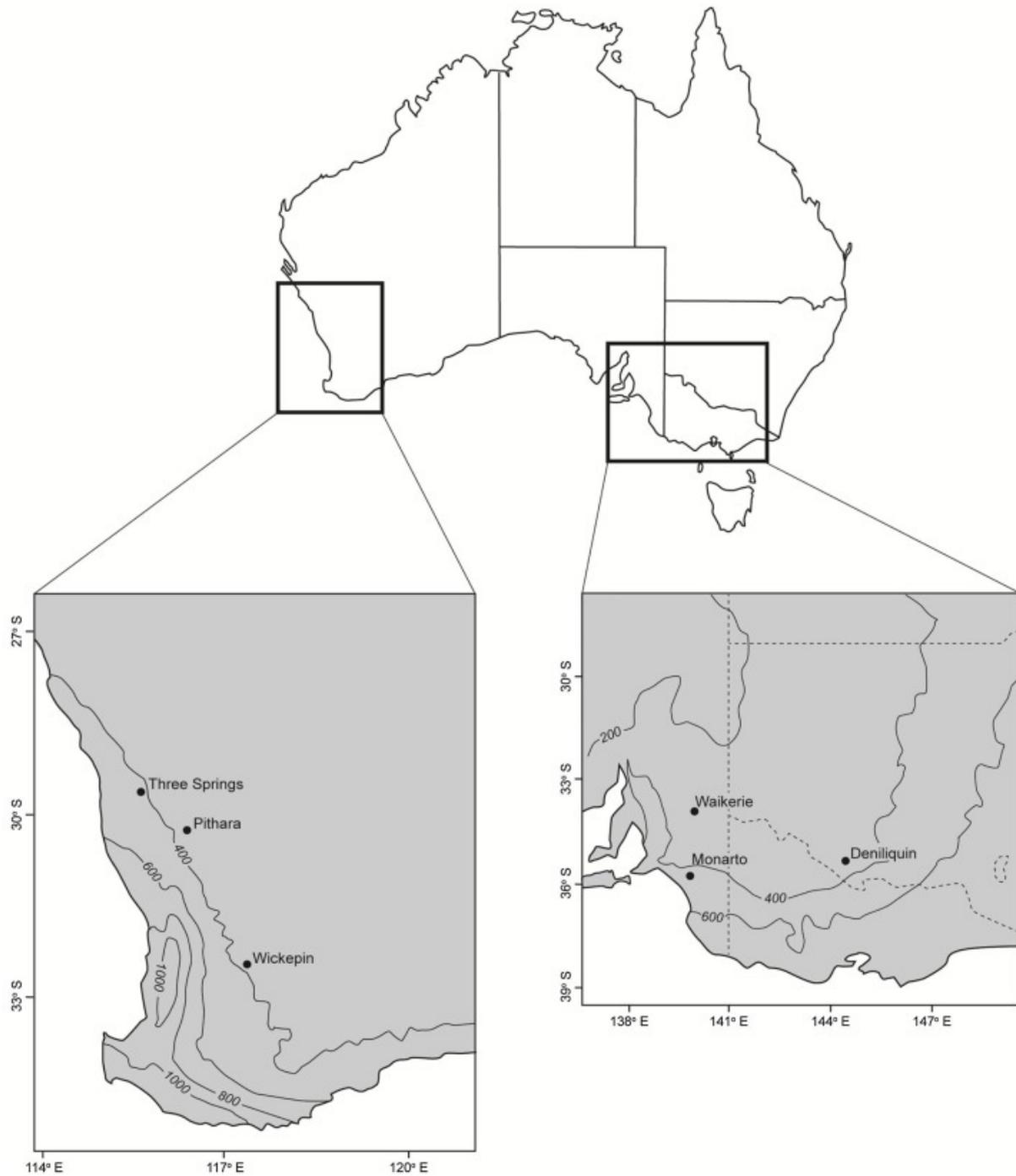


Fig. 2. Biomass data for each site. a) The mean percentage (%) of each biomass component at the four sites with above and below ground data and b) mean root to shoot ratio at each of the sites, and the mean when sites were combined. Bars are s.e.

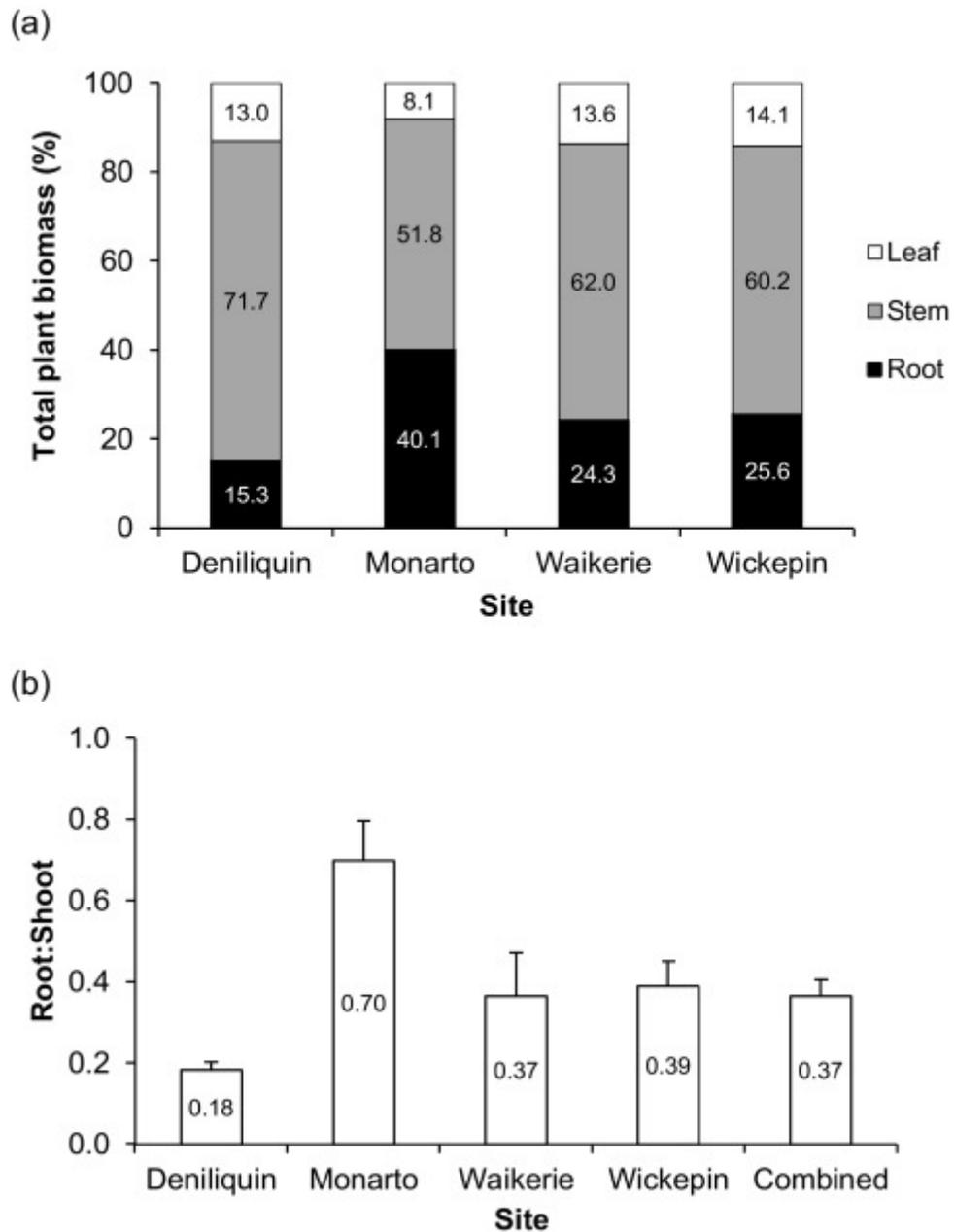


Fig. 3. Total dry biomass (TB) compared with the different plant growth estimators for all sites combined. a) height, b) crown diameter and c) crown volume index (CVI).

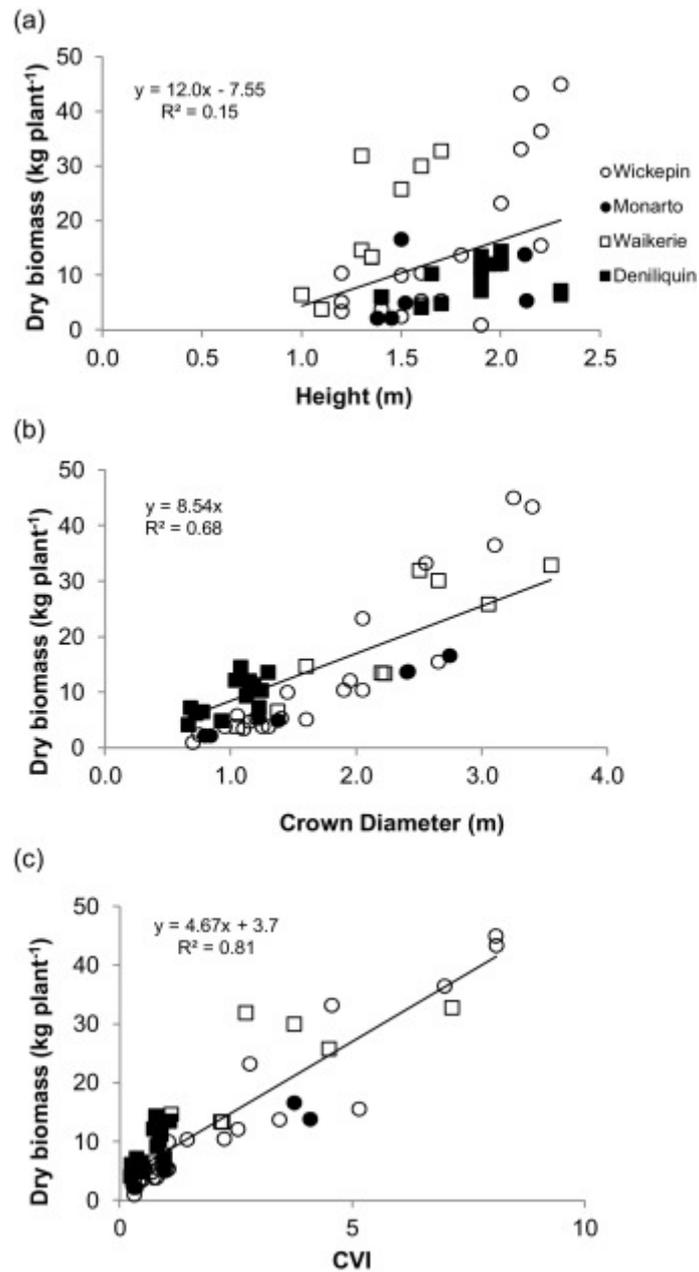


Fig. 4. Carbon sequestered over time at Wickepin for plots of two planting densities (LD: low density, 500 plants ha<sup>-1</sup>; HD: high density 2000 plants ha<sup>-1</sup>).

