Substantial Differences in Specificity of HIV-specific
Cytotoxic T Cells in Acute and Chronic HIV Infection

By Philip J.R. Goulder,*‡ Marcus A. Altfeld,* Eric S. Rosenberg,*
Thi Nguyen,* Yanhua Tang,* Robert L. Eldridge,* Marylyn M. Addo,*
Suqin He,* Joia S. Muckerjee,* Mary N. Phillips,* Michael Bunce,§
Spyros A. Kalams,* Rafick P. Sekaly,i Bruce D. Walker,*
and Christian Brander*

From the *Partners AIDS Research Center, Massachusetts General Hospital and Harvard Medical
School, Charlestown, Massachusetts 02129; the ‡Division of Infectious Diseases, The Children’s
Hospital, Boston, Massachusetts 02115; the §Oxford Transplant Centre, Churchill Hospital, Oxford
OX3 7LJ, United Kingdom; and the iLaboratoire d’Immunologie, Institut de Recherches Cliniques de
Montreal, Montreal, Quebec H2W 1R7, Canada

Abstract

Cytotoxic T lymphocytes (CTLs) play a vital part in controlling viral replication during hu-
man viral infections. Most studies in human infections have focused on CTL specificities in
chronic infection and few data exist regarding the specificity of the initial CTL response in-
duced in acute infection. In this study, HIV-1 infection in persons expressing human histo-
compatibility leukocyte antigen (HLA)-A*0201 was used as a means of addressing this issue. In
chronic infection, the dominant HLA-A*0201–restricted CTL response is directed towards the
epitope SLYNTVATL (“SL9”) in p17 Gag (residues 77–85). This epitope is targeted by 75%
of HLA-A*0201–positive adults, and the magnitude of this A*0201-SL9 response shows a
strong negative association with viral load in progressive infection. Despite using the highly
sensitive peptide–major histocompatibility complex tetramer and intracellular cytokine assays,
responses to the SL9 epitope were not detectable in any of 11 HLA-A*0201–positive subjects
with acute HIV-1 infection (P = 3 × 10^-6), even when assays were repeated using the SL9
peptide variant that was encoded by their autologous virus. In contrast, multiple responses
(median 3) to other epitopes were evident in 7 of the 11 A*0201–positive subjects. Longitudi-
nal study of two subjects confirmed that the A*0201-SL9 response emerged later than other
CTL responses, and after viral set point had been reached. Together, these data show that the
CTL responses that are present and that even may dominate in chronic infection may differ
substantially from those that constitute the initial antiviral CTL response. This finding is an
important consideration in vaccine design and in the evaluation of vaccine candidates.

Key words: acute • chronic • HIV infection • immunodominance • epitope targeting

Introduction

Virus-specific CTLs play a critical role in the control of vi-
ral infections (1–9). Studies in the lymphocytic choriomen-
ingitis virus (LCMV) mouse model have most clearly
shown that the early antiviral cellular immunity that is gen-
erated strongly influences the subsequent course and out-
come from the infection (10). Similar conclusions may be
drawn from investigations of human viral infections (3–5,
11–14). This implies that the immune responses that are ob-
served in the chronic phase of persistent virus infections
may largely be a consequence of what has occurred ini-
tially, and that perhaps the most important antiviral im-
mune responses to understand are those responsible for
early control of viremia in acute infection. Recent studies
in the simian immunodeficiency virus (SIV) macaque

P.J.R. Goulder and M.A. Altfeld contributed equally to this work.
Address correspondence to P.J.R. Goulder, Partners AIDS Research
Center, Massachusetts General Hospital, 13th St., Bldg. 149, Rm. 5218,
Charlestown, MA 02129. Phone: 617-726-5787; Fax: 617-726-5411;
E-mail: goulder@helix.mgh.harvard.edu

1Abbreviations used in this paper: BCL, B lymphoblastoid cell line; Elispot,
enzyme-linked immunospot; HAART, highly active antiretroviral therapy;
ICS, intracellular cytokine staining; SIV, simian immunodeficiency virus.

Published January 8, 2001
model strongly support this hypothesis (15). In the design and testing of vaccines, it is vital to know whether the CTL specificities differ significantly between acute and chronic infection. If differences do exist, it is necessary to determine which responses should optimally be generated by a candidate vaccine.

HIV-1 infection offers an excellent system in which to compare immune responses in acute and chronic infection. In this infection, CTLs are centrally involved both in the initial containment of viremia in acute infection and in limiting viral replication in chronic infection (16–30). However, evidence accumulating both from animal models and human viral infections indicate that significant differences exist between the effectiveness of CTLs of different specificities in controlling viremia (28–39). A recent study in the SIV macaque model of AIDS virus infection showed that initial immune pressure was directed mainly through an epitope in Tat; whereas one in Gag did not appear to be under strong immune selection pressure (15). Thus, from these data, it is likely that CTLs of different specificities may show dramatic differences in the contribution that each may make to the successful control of viremia.

The best studied HIV-specific CTL response is the dominant HLA-A*0201-restricted specificity that is directed towards an epitope in p17 Gag, SLYNTVATL (residues 77–85; references 22 and 40–46), referred to as “SL9.” In four independent studies of A*0201-positive adults with chronic HIV infection, 75% of persons studied had a detectable response towards this epitope (22, 44–46). Using SL9-A*0201 peptide–MHC tetrameric complexes, a strong negative association was shown between levels of SL9-A*0201 CTLs and viral load in A*0201-positive adults with chronic HIV infection (22). This widely quoted landmark study has provided strong indirect evidence that A*0201-SL9-specific CTLs are effective in controlling HIV replication in chronic infection. This has led to the suggestion that an effective vaccine should induce this response.

To determine whether A*0201-SL9-specific CTLs play an important role in the initial control of viremia in acute infection, the HIV-specific CTL responses were characterized in 11 subjects with A*0201 who were enrolled early in the course of HIV infection, in 8 cases before seroconversion. Although CTL responses were observed in the majority of these subjects, none made a detectable response to the A*0201-SL9 epitope.

Materials and Methods

Subjects Studied. The 11 A*0201-positive subjects studied in early HIV infection were recruited from persons presenting to the Massachusetts General Hospital between 1997 and 1999 who were diagnosed with acute HIV infection. The relevant clinical data for these subjects are described in Table I. All but three of the subjects were enrolled before seroconversion: three subjects (AC14, AC23, and AC32) were enrolled within 180 d of seroconversion, as defined by an adapted ELISA assay for HIV Ab (47), and one subject (AC29) was included using clinical criteria that supported a diagnosis of HIV infection within the previous 3 mo. These included a known high-risk exposure to HIV infection, by history a clinical syndrome consistent with acute HIV infection, and a viral load at the time of presentation of >750,000 RNA copies/ml plasma that would be highly unusual as a steady-state viral load in chronic infection (14, 48). All 11 subjects were initially studied before initiation of highly active antiretroviral therapy (HAART) as detailed. An additional A*0201-positive subject PI004 whose date of acquiring HIV infection was less certain, and who was never treated with HAART, was also studied as described below. The first sample available for study on PI004 was 7 wk after the first positive HIV Ab test on this subject. An HIV Ab test 21 wk previous to this had been negative.

HLA Class I Tissue Typing and HLA-A2 Subtyping. HLA class I typing and A2 subtyping were performed by sequence-specific primer (SSP)-PCR (49). Only subjects with A*0201 were included in the study.

Peptides. PBMCs in each of the 11 subjects with early HIV infection were screened for recognition in enzyme-linked immunosorbent (Elispot) assays of epitopes within p17 Gag, p24 Gag, Nef, RT, gp41, gp120, Tat, and Rev using overlapping peptides 12–20 amino acids in length that overlapped by 10 amino acids. Overall, 290 overlapping peptides were used to span these 8 proteins. In addition, from a total of 130 published optimal epitope peptides (50), those that were presented by HLA class I molecules expressed by each subject studied (median 24 peptides) were tested for recognition in Elispot assays. The sequences for the peptides corresponded to the B clade SF2 sequence. Gag, Nef, RT, and gp120 peptides were provided by the National Institute for Biological Standards and Control Centralized Facility for AIDS Reagents, supported by European Union Program EVA and the UK Medical Research Council; 12 additional overlapping p17 Gag peptides, gp41, Tat, and Rev peptides were synthesized commercially (Research Genetics) or at the Massachusetts General Hospital Peptide Synthesis Core Facility.

Elispot Assays. Fresh PBMCs were plated in 96-well polystyrene plates (Millipore) that had been precoated with 0.5 μg/ml anti-IFN-γ mAb, 1-DIK (Mabtech). The peptides were added in a volume of 20 μl and PBMCs were added at 100,000 cells/well in a volume of 180 μl RPMI 1640 (Sigma-Aldrich), 10% FCS [Sigma-Aldrich], and 10 mM Hepes buffer [Sigma-Aldrich] with antibiotics [2 μM 1-glutamine, 50 μ/ml penicillin-streptomycin]). The end concentration of the peptides was 10 μM. The plates were incubated overnight at 37°C, 5% CO₂, and developed as described previously (51, 52). The number of specific T cells was calculated by subtracting the negative control values. The background was <20/10⁶ PBMCs (2 spots/well at 100,000 PBMCs/well) in all cases. Responses of >60 IFN-γ spot-forming cells/10⁶ PBMCs were therefore significant positive responses, and these were reconfirmed in intracellular IFN-γ staining assays (see below). Wells that contained >30 spots were not used for accurate quantification. Assays were repeated using lower input numbers of cells as necessary and in duplicate in order to quantitate responses to individual peptides more accurately.

Intracellular IFN-γ Staining. Intracellular cytokine staining (ICS) assays were performed as described elsewhere (53–55). In brief, 0.2–1.0 × 10⁶ PBMCs were incubated with 4 μM peptide and 1 μg/ml each of the mAbs anti-CD28 and anti-CD49d (Becton Dickinson) at 37°C, 5% CO₂ for 1 h, before the addition of 10 μg/ml of Brefeldin A (Sigma-Aldrich). After a further 6 h incubation at 37°C, 5% CO₂, the cells were placed at 4°C overnight. PBMCs were then washed and stained with surface Abs anti-CD8 and anti-CD3 (Becton Dickinson) at 4°C for 20 min.
PBMCs that were also stained with tetramers were incubated with the tetramer at 4°C for 30 min before the addition of the surface Abs. After washing, the PBMCs were then fixed and permeabilized (Caltag) and anti–IFN-γ mAb was added (Becton Dickinson). Cells were then washed and analyzed. Quadrant boundaries for IFN-γ staining were established by exclusion of >99.97% of control CD8+ T cells.

**Peptide–MHC Tetramer Assays.** Peptide–MHC tetramers were synthesized as described previously (42, 56). The tetramer used in these studies was the HLA-A*0201–SLYNTVATL complex. HLA heavy chain was expressed in *Escherichia coli* with an engineered COOH-terminal signal sequence containing a biotinylation site for the enzyme BirA. After refolding of heavy chain, βµm, and peptide, the complex was biotinylated by BirA (AviTag) in the presence of ATP-Mg2+ (Sigma-Aldrich). After purification by gel filtration and anion exchange chromatography, tetramer formation was induced by the addition of streptavadin. Use of PE-labeled streptavidin enabled antigen-specific cells to be visualized by flow cytometry.

Staining of lymphocytes was performed by incubating 500,000 PBMCs for 30 min at 4°C with the appropriate tetramer at 0.5 mg/ml of tetramer, then for a further 20 min with saturating amounts of peridinin chlorophyll protein (PerCP)-conjugated anti–CD8 mAb and allophycocyanin (APC)-conjugated anti–CD4 mAb (Becton Dickinson). Stained samples were analyzed on a FACSCalibur™ flow cytometer using CELLQuest™ software (Becton Dickinson). Control samples for the tetramer staining were PBMCs from HLA-mismatched HIV-infected persons. Quadrant boundaries for tetramer staining were established by exclusion of >99.97% of control CD8+ T cells.

**Generation of CTL Clones, Precursor Frequency Assays.** CTL clones were generated using methods described previously (57). In brief, PBMCs were plated out in 96-well plates at limiting dilution (30 cells/well down to 1 cell/well) and cultured with irradiated allogeneic feeder PBMCs at 50,000 cells/well in a final volume per well of 200 µl of R10. The anti–CD3 mAb, 12F6, was added at 10 µg/ml. On day 5 and once weekly thereafter, the medium was changed with R10 medium containing 50 U/ml of rIL-2 (provided by Dr. M. Gately, Hoffmann-La Roche, Nutley, NJ). Wells were screened for specific recognition of HLA-matched, peptide-pulsed, 51Cr (New England Nuclear)-labeled EBV-transformed B lymphoblastoid cell line (BCL) target cells after 21–28 d in culture. Wells showing high specific recognition of the relevant peptide were then transferred to 24-well plates and restimulated as above, except 106 feeders were added to each well and rIL-2 was added on day 0. Expanded wells were then restested for lytic activity from 14 d of culture onwards, and maintained in culture by monthly restimulations as described (57).

**Cr Release Assays.** BCL target cells were labeled with 51Cr by incubation of pelleted BCL with 50 µCi of Na2CrO4 (New England Nuclear) for 1 h at 37°C, 5% CO2. Targets were washed three times and then incubated with peptide dilutions in the peptide titration assays for a further 90 min, before addition of effectors. The supernatants were harvested after a further 4–6 h of incubation at 37°C, 5% CO2 (58).

**Sequencing of Viral DNA and Sequence Analyses.** Genomic DNA was extracted from frozen PBMC pellets (3 × 106 cells) using the Puregene™ DNA isolation kit (Gentra). HIV Gag sequences were amplified by nested PCR using inner and outer primer sets and PCR conditions as described previously (44). For sequencing cloned viral sequences, 2 µl of the gel-purified PCR product was used for ligation of the Gag sequences into the Topo2 cloning plasmid according to the manufacturer’s recom
mendation (Invitrogen) and plasmid DNA was obtained after transfection of E. coli cells and DNA purification using the QIAGEN Turbo DNA purification kit. Inserted Gag sequences were determined from both directions on an ABI 377 sequencer using primer sequences located in the plasmid sequence on both sides of the insert (Topo2 cloning kit; Invitrogen). Sequence analysis was performed using the sequencer software version 3.1.1. BLAST was used to compare sequences from each of the study subjects with each other and sequences in the viral subsection of GenBank to screen for potential cross-contamination. Signature analysis was done to verify viral sequence identity and phylogenetic tree analyses using the Neighbor TreeMaker program, available at the Los Alamos HIV Database web site (http://hiv-web.lanl.gov/CONTAM/TreeMaker/TreeMaker.html), revealed unique signature sequences for the individuals. These sequence data are available from Genbank/EMBL/DDBJ under accession nos. AF281678 and AF281801.

Results

Recognition of the HLA-A*0201-restricted p17 Gag Epitope in Acute Infection. To characterize the HIV-specific CTL response in acute infection for each subject enrolled, IFN-γ responses to epitopes within p17 Gag, p24 Gag, Nef, RT, gp41, gp120, Tat, and Rev were screened in Elispot assays using panels of overlapping 15–20-mer peptides that overlapped by 10 amino acids to span each protein. The approach that was used is illustrated for one subject in Fig. 1 A. In addition, individual peptides previously defined as optimal epitopes corresponding to the HLA class I alleles expressed by each subject were tested for recognition. Thus, for each of the 11 A*0201-positive subjects studied with early HIV infection, 290 overlapping 15–20-mer peptides and between 11 and 29 (median of 24) optimal epitope peptides were used to characterize the CTL response. Overall, 78 different optimal epitope peptides were used in the studies of 11 A*0201-positive subjects with early infection.

None of the 11 subjects studied in this way had detectable CTL activity towards the A*0201-SL9 epitope (Fig. 1 B), even when responses were evaluated at several time points in the first 12 mo after presentation. In comparison, responses towards multiple epitopes other than SL9 (range 1–7, median 3) were observed in 7 of these A*0201-positive 11 subjects during this first year after the presumed time of infection (59). These data clearly contrast with the...
frequent detection of A*0201-SL9–specific responses (in 75% of A*0201-positive adults with chronic infection; n = 72) that has been described in four independent studies (22, 44–46) ($\chi^2 = 20.9, P = 2 \times 10^{-6}$). Even restricting the comparison to the seven A*0201-positive subjects who had no detectable SL9 response but who showed evidence of other HIV-specific CTL activity, the absence of an A*0201-SL9 response remains strongly significant ($P < 2 \times 10^{-4}$, Fisher’s exact test).

The Elispot assay has a high sensitivity but this does not quite reach the sensitivity of the flow cytometric assays to detect antigen-specific CD8$^+$ T cells either by intracellular IFN-γ staining after peptide stimulation, or by peptide–MHC class I tetrameric complexes (53, 60). In addition, tetramer assays have revealed the presence of phenotypically silent antigen-specific T cells in both murine and human viral infections (3, 38, 39). These assays were therefore used to determine whether any low-frequency or phenotypically silent A*0201–SL9–specific CD8$^+$ T cells could be detected using these highly sensitive assays. All A*0201-positive subjects either showed responses to an A*0201–EBV peptide, GLCTLVAML (61) and/or an A*0201 CMV peptide, NLVPMVATV (62), and/or a positive control HIV peptide in the ICS assay, but none recognized the A*0201-SL9 epitope (Fig. 2, and data not shown). The A*0201-SL9 tetramer and the SL9 peptide in the ICS assay demonstrated responses to this epitope in an A*0201-positive subject 9354 with chronic infection. Thus, no phenotypically silent A*0201-SL9 tetramer-binding cells were detectable in the subjects studied.

To reconfirm that no low-frequency A*0201-SL9–specific CTLs were detectable by any of the most sensitive assays, in two subjects a further method involving stimulation of PBMCs with SL9 peptide before culture in IL-7–containing medium (63) that has been successfully used to detect low-frequency CTL responses (64) was employed. However, this method was only successful in generated SL9-specific responses in A*0201-positive subjects with chronic infection, and not in those with acute infection (data not shown).

**Variant Epitope Sequence Encoded by Transmitted Virus Does Not Explain the Absence of an SL9-specific CTL Response in Acute Infection.** To investigate why no responses were generated towards the A*0201-SL9 epitope in acutely infected subjects with A*0201, autologous virus was sequenced to determine whether mutated epitope sequences had been transmitted. It was hypothesized that this could explain the absence of an SL9-specific response if the autol-

![Figure 2](https://jem.rupress.org/content/jem/8/1/185/F2.large.jpg)

**Figure 2.** Recognition of SLYNTVATL only in A*0201-positive subjects in chronic infection. Intracellular staining using SL9 and a positive control HIV peptide epitope and A*0201-SL9 tetramer staining of PBMCs from chronically infected subject 9354 (HLA-A*0201/3 B7/35 Cw4/7) and 2 of the 11 A*0201-positive subjects studied in acute infection (expressed as percentage of CD8$^+$ T cells). Controls: percentage of CD8$^+$ T cells showing intracellular IFN-γ staining after incubation with no peptide was, respectively, 0.00% (subject 9354), 0.03% (AC29), and 0.02% (AC13; data not shown). The HIV peptides used as positive controls had previously been established as recognized by these subjects: B7 Gag TL9, TPQDLNTML (p24 Gag); B8 Nef FL8, FLKEKGGL; and B14 gp41 EL9, ERYLKDQQL (reference 50).
ogous SL9–epitope sequence differed significantly from the consensus B clade sequence. In 10 of the 11 A*0201-positive subjects enrolled, the autologous gag sequences encoding the p17 Gag SL9 epitope were determined (Table II). Only sequences encoding full-length p17 and p24 Gag were included in the analysis. In all subjects, the predominant autologous sequence encoded either the B clade consensus sequence SLYNTVATL or variants that occur very frequently in Los Alamos database sequences (50). Even when these A*0201-positive acutely infected subjects were tested for recognition of autologous SL9 variants, still none recognized his autologous epitope. (Fig. 3, A and B).

To demonstrate further that variants of SL9 should be able to induce CTL responses, CTL clones specific for this response were generated in chronically infected subjects with A*0201 and the SL9 variants occurring most frequently in published sequences were tested for recognition. The five SL9 variants that were tested, together with the clade B consensus sequence SLYNTVATL, account for 89% of the 272 B clade Los Alamos database sequences originating in the USA (50). Many distinct patterns of variant recognition were observed, in several cases showing that SL9 variants can be recognized as well or better than the consensus sequence (Fig. 3, C–H). Although a formal comparison of the ability of SL9 and the different SL9 variants to induce CTL responses was not undertaken, these data together imply that transmission of virus that encodes the commonly occurring SL9 variants (illustrated in Fig. 3) is not a barrier to the generation of CTL responses towards this epitope.

It should be noted that although the SL9 (SLYNTVATL) sequence is considered the consensus clade B sequence, as many sequences in the database contain SL9 variants as contain the consensus sequence within this highly variable region. Thus, it is highly unlikely that the 72 A*0201 subjects studied in chronic infection were all infected with virus encoding the SL9 consensus sequence. Indeed, in one of these studies (45), only 3/11 A*0201-negative chronically infected control subjects had virus encoding this consensus SL9 sequence, a virtually identical proportion to that observed in the acutely infected A*0201-positive subjects described here. However, even if we restrict our analysis to the subjects with A*0201 whose autologous virus encoded the so-called consensus SL9 epitope, the absence of an SL9 response in acute infection still differs significantly from the chronically infected A*0201-positive subjects, 75% of whom made a response to SL9 (P < 0.02, Fisher’s exact test).

**Table II.** SLYNTVATL Epitope Variants Encoded in 10 of the 11 Subjects with Early/Acute HIV Infection

<table>
<thead>
<tr>
<th>Subject</th>
<th>Date*</th>
<th>Clones analyzed</th>
<th>S</th>
<th>L</th>
<th>Y</th>
<th>N</th>
<th>T</th>
<th>V</th>
<th>A</th>
<th>T</th>
<th>L</th>
</tr>
</thead>
<tbody>
<tr>
<td>AC23</td>
<td>11/10/89</td>
<td>8/8</td>
<td></td>
<td></td>
<td>F</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>AC29</td>
<td>03/09/99</td>
<td>11/11</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>AC04</td>
<td>10/17/97</td>
<td>13/13</td>
<td></td>
<td></td>
<td>F</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>AC01</td>
<td>01/31/97</td>
<td>11/12</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>I</td>
<td>V</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>1/12</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>P</td>
<td>I</td>
<td>V</td>
<td></td>
<td></td>
</tr>
<tr>
<td>AC03</td>
<td>09/01/97</td>
<td>13/15</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>A</td>
<td></td>
</tr>
<tr>
<td></td>
<td>1/15</td>
<td></td>
<td></td>
<td></td>
<td>I</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>AC14</td>
<td>07/27/99</td>
<td>13/13</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>I</td>
<td>V</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>AC13</td>
<td>06/25/98</td>
<td>11/11</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>I</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>AC22</td>
<td>10/30/99</td>
<td>12/14</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>V</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>2/14</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>T</td>
<td>V</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>AC26</td>
<td>01/21/99</td>
<td>14/14</td>
<td></td>
<td></td>
<td>F</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>AC32</td>
<td>05/26/99</td>
<td>6/6</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>I</td>
<td>V</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*Viral sequence obtained from samples taken prior to seroconversion from all subjects except AC14, AC23, AC29, and AC32.
The frequency of A*0201-SL9–specific CTLs by tetramer assay was 0.07% of PBMCs, compared with 0.03% of PBMCs in the Elispot assay (data not shown). The magnitude of this response increased once it had developed, and in the absence of any substantial reduction in the magnitude of the other five responses.

Although the pattern of responses in subject AC13 over time is of interest, these data are complicated by the fact that AC13 had previously received HAART from 2 wk after presentation and had also undergone two treatment interruptions. At the time the A*0201 Gag response emerged, AC13 was not on antiretroviral therapy and had detectable viremia. In an additional A*0201-positive subject PI004, recruited from a separate small cohort of acutely infected persons from Montreal, who was untreated throughout the time of study, a similar phenomenon of the late appearance of SL9-specific CTLs in the HIV-specific response was observed. This response was detectable 5 mo later than three responses that were present from the first time point available for study onwards (Fig. 4 B). Of note, in both AC13 and PI004, initial control of viremia in acute infection had been achieved in the absence of an A*0201-SL9–specific response. In AC13, the viral load had fallen from 730,000 to 5,000 HIV-1 RNA copies/ml plasma before the initiation of HAART. In PI004, the viral load for the first four time points shown, before the appearance of
the A2-Gag–specific CTL response, was between 50 and 400 RNA copies/ml plasma. Thus, the A*0201-SL9–specific response is not required for the initial control of viremia in A*0201-positive subjects.

Discussion

These studies show that the HIV-specific CTL epitopes that may dominate in chronic infection may play no part in the critical antiviral CTL response associated with initial clearance of virus in acute infection. Not one of 11 A*0201-positive subjects with early HIV infection had detectable CTL responses towards the p17 Gag SL9 epitope that is recognized in 75% of A*0201-positive adults with chronic infection ($P = 2 \times 10^{-6}$; references 22 and 44–46). This absence of a response to SL9 in acute infection was especially striking in the seven A*0201 subjects who showed readily detectable responses to multiple other HIV-specific epitopes ($P < 0.0002$). Finally, the delayed appearance of an A2-restricted CTL response towards the SL9 epitope well after responses were detectable to other epitopes was confirmed in longitudinal studies of two subjects.

In addition to the consensus SL9 sequence, autologous variants were also tested, and there was also no recognition of either autologous or consensus SL9 sequence. This was important because previous studies in EBV have suggested viral evolution over time to mutate immunodominant epitopes (65). We have shown previously that mutations within and adjacent to the SL9 epitope do not alter processing to affect recognition of the SL9 variant epitope (66). Thus, there is no clear explanation from the viral sequence data presented for the absence of the A*0201-SL9 response in acute infection.

To determine whether A*0201-SL9–specific CTLs were present in early infection that were functionally inert (3, 38, 39), tetramer assays showed conclusively that no phenotypically silent antigen–specific CD8+ T cells could be detectable in the A*0201 subjects studied in early HIV infection. In longitudinal studies of subject AC13, A*0201-
SL9 tetramer-binding cells were detectable only when SL9–specific responses in the Elispot and intracellular cytokine assays became evident.

These data are critical in relation to vaccine design, as they raise the question of whether the responses that dominate in chronic infection are in fact important in control of HIV, as was believed previously (22). It is clear from subjects AC13 and PI004 that initial control of viremia in these two subjects was achieved without any contribution from the A*0201–SL9 response. Thus, an A*0201–SL9 response is evidently not required for effective control of acute viremia. It is possible that the A*0201–SL9 CTL response in chronic infection may only be associated with control of viremia indirectly, through the action of CTLs of different HIV specificities, or via HIV-specific T helper responses. Resolving this important question will require further detailed work to understand the HIV-specific T response. Resolving this important question will require additional work to understand the HIV-specific T helper responses. Resolving this important question will require additional work to understand the HIV-specific T helper responses.

The factors likely to contribute to the dominance of the A*0201–SL9 response in natural HIV infection indicate that the entire HIV-specific CTL response cannot be estimated adequately by focusing on this single specificity.

It remains unresolved why the A*0201–SL9 responses arise late. The factors likely to contribute to the dominance of CTL responses (68) in this case include the binding affinity of SL9 to HLA-A*0201 (69, 70), the efficiency of processing of the SL9 epitope (41, 66, 71), and the presence of CTL escape (44, 72) in subjects that were previously immunodominant.

It is clear from binding studies already performed that SLYNTVATL is not a strong binder to A*0201 (45, 73–76), although all the SL9 variants tested do indeed bind. The gp41–A*0201 epitope, SLLNATAIAV (SV10), that dominates the early CTL response in subject AC13 (Fig. 4A) in fact in recent studies proved to be the strongest binder to A*0201 of all HIV-1 peptides tested that had been selected on the basis of motif (76). Thus, the relatively weak binding of SL9 to A*0201 may contribute to the relatively poor immunogenicity of this epitope.

A second factor likely to contribute to the dominance of the early CTL response would be the efficiency with which individual epitopes are processed. It might be anticipated that epitopes derived from the regulatory and accessory proteins such as Tat, Rev, and Nef, expressed in abundance on the surface of infected cells early in the viral life cycle (77), might dominate the acute antiviral CTL response (15). However, in the cohort of 19 subjects with acute HIV infection being studied in Boston, Gag–specific epitopes were in fact targeted more frequently than Nef–specific at preseroconversion time points (33 vs. 19%; reference 59). Furthermore, no Nef–, Tat–, or Rev–specific A*0201–restricted responses were detectable in any of the 11 A*0201 subjects studied here in early HIV–1 infection (data not shown). Thus, low expression of Gag (relative to Tat, Rev, or Nef) in acute infection does not appear to be an explanation for the absence of A*0201–SL9 responses at this time.

A further possible explanation for the late appearance of the A*0201–SL9 response in the course of HIV–1 infection would be that CTL escape occurring in the dominant epitopes would enable previously subdominant epitopes to become immunodominant over time (72). The limited data available from subject PI004 (Fig. 4B) do appear to indicate that a substantial reduction in the dominant HLA-B57–restricted responses towards p24 Gag epitope TSLQEQIGW (34, 50) has occurred immediately before appearance of the A*0201–SL9 and several other responses that previously were undetectable. In AC13, the data are more difficult to interpret, as this subject was twice treated with HAART, but the appearance is more suggestive in this case that the A*0201–SL9 response emerges at a time when the CTL response as a whole is increasing in magnitude. These and other subjects who show the late appearance of particular epitopes clearly warrant further study in this regard.

It is important to note that, although SL9 is the dominant A*0201–restricted response in chronic infection, it is not commonly the dominant HIV–specific CTL response overall (34, 78–80). Thus, it is not unexpected to see that SL9 is subdominant to B14– and B57–restricted responses in subjects AC13 and PI004, and that the SL9 response does not increase to become dominant to these other responses.

The closest precedent for this phenomenon may be in human EBV infection, in which lytic as opposed to latent antigen–specific responses dominate in acute infection (2, 81). However, this difference corresponds to the different biological phases of EBV infection. In HIV infection, there is no indication that Gag is not expressed in acute infection; in fact, in 5/7 HLA–A3–positive subjects studied at preseroconversion, p17 Gag–specific HLA–A3–restricted activity was a major component part of the initial anti–HIV immune response (59). There are indications, both in studies of SIV (15) and HIV (59, 82), and also in hepatitis C virus infection (3, 83), that many clear-cut differences in the specificities of the CTL response in acute and chronic viral infections may exist. However, these data presented with respect to the A*0201–SL9 specificity are apparently the first unequivocal demonstration of this phenomenon.

In conclusion, these data show that the HIV–specific CTL responses that are present in acute infection may differ substantially from those that are frequently detectable in chronic infection. The best–studied response in chronic
HIV infection is seen to play little or no part in the antiviral immune response in acute infection that is so critical in determining the ultimate outcome from infection. Further studies are needed to assess the importance of the CTL responses that are frequently detectable in subjects with chronic infection, to determine whether these, or the responses present in acute infection, or both, are effective responses that should be incorporated into HIV vaccine design.

We thank Paul Klenerman and David Watkins for helpful comments and discussion of the manuscript.

This work was supported by grants to P.J.R. Goulder from the Elizabeth Glaser Pediatric AIDS Foundation, the Medical Research Council (UK; grant G108274), and the National Institutes of Health (grant AI46995); to M.A. Altfield through the German Academic Exchange Foundation; to E.S. Rosenberg through the Doris Duke Charitable Foundation and the National Institutes of Health (grant AI01541); to P.J.R. Goulder through the German Research Foundation; to S.A. Kalams through the National Institutes of Health (grant AI30914); and to B.D. Walker through the National Institutes of Health (grants AI28568 and AI30914) and the Doris Duke Charitable Foundation. P.J.R. Goulder is an Elizabeth Glaser Pediatric AIDS Foundation Professor.

Walker is a Doris Duke Distinguished Clinical Science Professor.

Duke Charitable Foundation. P.J.R. Goulder is an Elizabeth Glaser Pediatric AIDS Foundation, the Medical Research Council (UK; grant G108274), and the National Institutes of Health (grant AI46995); to M.A. Altfeld through the German Academic Exchange Foundation; to E.S. Rosenberg through the Doris Duke Charitable Foundation and the National Institutes of Health (grant AI01541); to M.M. Addo through the German Research Foundation; to E.S. Rosenberg through the Doris Duke Charitable Foundation and the National Institutes of Health (grant AI30914) and the Doris Duke Charitable Foundation.

Submitted: 23 June 2000
Revised: 12 October 2000
Accepted: 28 November 2000

References

Dramatic rise in plasma viremia after CD8+ lymphocyte response

Evans, D.T., D.H. O’Connor, P. Jing, J.L. Dzuris, J. Sidney,

Borrow, P., H. Lewicki, X. Wei, M.S. Horwitz, N. Peffer,


Koenig, S., A.J. Conley, Y.A. Brewah, G.M. Jones, S. Leath,


Kaslow, R.A., M. Carrington, R. Apple, L. Park, A. Munoz,


